

Single-cell lineage tracing with a focus on cancer metastasis using macsGESTALT

Pinello Lab Journal Club

Friday | June 25th, 2021



Overview of today's presentation

Paper we are covering today:

Cancer Cell

Article

Single-cell lineage tracing of metastatic cancer reveals selection of hybrid EMT states

First we will cover some history, background, motivation, and context:

RESEARCH ARTICLE

CELL LINEAGE TRACING

Whole-organism lineage tracing by combinatorial and cumulative genome editing

Aaron McKenna,^{1*} Gregory M. Findlay,^{1*} James A. Gagnon,^{2*} Marshall S. Horowitz,³ Alexander F. Schier,^{2,4,5,6,†} Jay Shendure^{1,7,†}

Developmental Cell

CellPress

Review

Next-Generation Lineage Tracing and Fate Mapping to Interrogate Development

Science

RESEARCH ARTICLES

Cite as: Weinreb *et al.*, *Science* 10.1126/science.aaw3381 (2020).

Lineage tracing on transcriptional landscapes links state to fate during differentiation

Rigoberto Fraticelli^{2,3*}, Fernando D. Camargo^{2,3†}, Allon M. Klein^{1†}

SINGLE-CELL OMICS

Building a lineage from single cells: genetic techniques for cell lineage tracking

Mollie B. Woodworth¹⁻³, Kelly M. Girsakis¹⁻³ and Christopher A. Walsh¹⁻³

Article

Cell

Lineage Tracing in Humans Enabled by Mitochondrial Mutations and Single-Cell Genomics

Leif S. Ludwig,^{1,2,15*} Caleb A. Lareau,^{1,2,3,4,15} Jacob C. Ulirsch,^{1,2,4,15} Elena Christ Karin Pelka,^{1,6,7} Will Ge,¹ Yaara Oren,^{1,8} Alison Brack,¹ Travis Law,¹ Christopher Genevieve M. Boland,^{5,10} Nir Hacohen,^{1,6,7} Orit Rozenblatt-Rosen,¹ Martin J. Ary Aviv Regev,^{1,13,*} and Vijay G. Sankaran^{1,2,14,16,*}

ARTICLE

<https://doi.org/10.1038/s41586-018-0744-4>

Single-cell mapping of lineage and identity in direct reprogramming

Brent A. Biddy^{1,2,3}, Wenjun Kong^{1,2,3}, Kenji Kamimoto^{1,2,3}, Chuner Guo^{1,2,3}, Sarah E. Waye^{1,2,3}, Tao Sun^{1,2,3,4} & Samantha A. Morris^{1,2,3*}

Lineage tracing meets single-cell omics: opportunities and challenges

Daniel E. Wagner^{1,2,3} and Allon M. Klein^{1,3}

Lineage reconstruction from clonal correlations

Weinreb^a and Allon M. Klein^{a,1}

Overview of today's presentation

Access all sides with links to papers and notes:



The screenshot shows a GitHub Wiki page for the repository 'mvinyard / vintools'. The page title is 'Pinello Lab Journal Club: June 25th, 2021', edited by Michael Vinyard. The main content is a featured paper titled 'Single-cell lineage tracing of metastatic cancer reveals selection of hybrid EMT states' from the journal 'Cancer Cell'. The paper's graphical abstract shows a mouse model where cancer cells are injected into the pancreas, followed by lineage recording and reconstruction. The authors listed are Kamen P. Simeonov, China N. Byrns, Megan L. Clark, Jay Shendure, Aaron McKenna, and Christopher J. Lengner. The page also includes a sidebar with navigation links to 'Home', 'google_vm', and the current page, along with a search bar and a 'Clone this wiki locally' button.

 **GitHub.com/mvinyard/vintools**



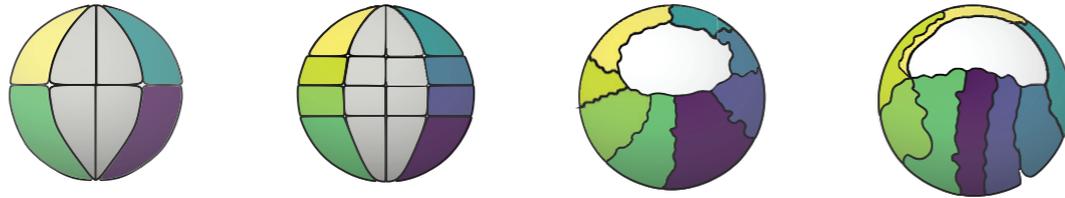
Overview of today's presentation

1. Brief history lesson and overview of tools / datasets / technologies available
2. Overview of GESTALT, the precursor technology to macsGESTALT
3. In-depth coverage of the macsGESTALT paper



Brief history of fate mapping and lineage tracing

Fate Map



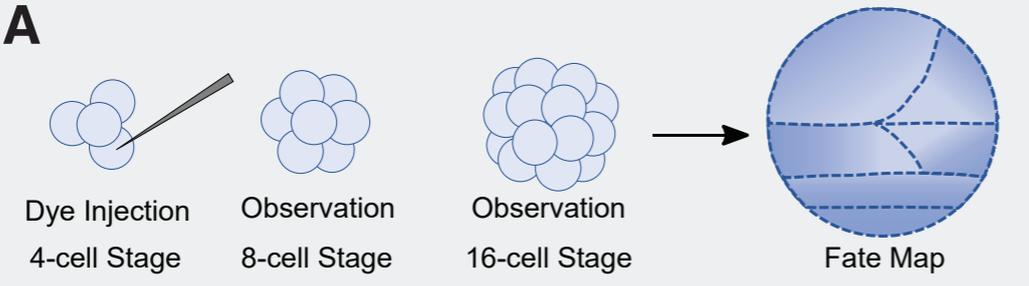
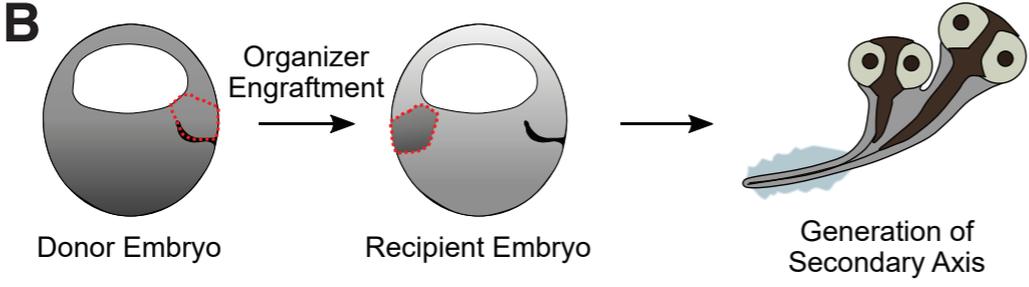
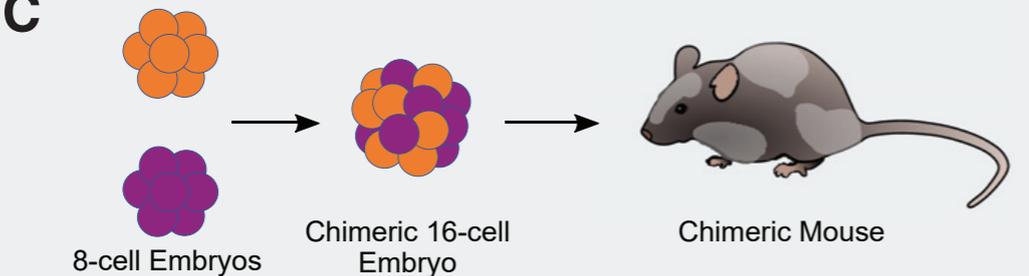
Fate maps = *schematics* of developmental potential

- Fate mapping and lineage tracing are related but **distinct**
- Lineage tracing identifies progeny from a given ancestor cell

Era	Year	Lineage Tracing Technique	Resolution	Scalability	Limitation	Technique & Citation
Observational Biology	1890s	A Dye Injection 4-cell Stage Observation 8-cell Stage Observation 16-cell Stage Fate Map	Single-cell limited by injection	10s of cells limited by observation	Observational data	Dye Injection and Time Lapse Conklin, 1905 Vogt, 1924
	1920-30s	B Organizer Engraftment Donor Embryo Recipient Embryo Generation of Secondary Axis	N/A	Tissues	Observational data	Organizer Grafts Spemann and Mangold, 1924 Wetzel, 1929
	1960s	C 8-cell Embryos Chimeric 16-cell Embryo Chimeric Mouse	N/A	Tissues	Only specific to embryo of origin	Chimera Generation Tarkowski, 1965 Mintz, 1965
Molecular Biology	1980s	D Retroviral Transduction Cellular Proliferation <i>in vivo</i> or <i>in vitro</i> Marker Recovery (i.e. β -gal)	Theoretically single clones	10s of cells limited by observation	Observational data	Retroviral Labelling Cepko <i>et al.</i> , 1987
	2000s	E Retroviral Transduction of Specialized Cre/lox Cassette Cre Activation Creates Alternative XFP Readout Lineage Determination	Theoretically single clones	100s of cells limited by observation	Observational data	Randomized Recombination Cassettes Livet <i>et al.</i> , 2007 Snippert <i>et al.</i> , 2010
	2010s	F Cas9 targeted editing of transgene or endogenous genomic DNA Accrual of Cas9-mediated scars on target sequences Clonal tracking of regeneration in caudal fin	Theoretically single clones	100s of cells limited by observation	Dataset limitation based on collection method	Cas9 Targeted Scar Accrual McKenna <i>et al.</i> , 2016 Junker <i>et al.</i> , 2017
Single-cell Biology	2010s-	G Retroviral Transduction with CellTags Rounds of CellTagging generate barcode combinations scRNA-seq reads include transcriptomic and lineage barcode data	Single-cell	1000s - 10,000s of cells	Resolved to clonal and sub-clonal populations	Retroviral mRNA Barcode Accrual Yao <i>et al.</i> , 2017 Biddy <i>et al.</i> , 2018 Weinreb <i>et al.</i> , 2020
	2010s-	H Cas9-targeted editing of genomic DNA sequence Accrual of Cas9-mediated scars on target sequences Generation of lineage trees based on mRNA barcode readout	Single-cell	1000s - 10,000s of cells	Information dropout due to Cas9 induced deletion of previous scars	Cas9 mRNA Scars Spanjaard <i>et al.</i> , 2018 Raj <i>et al.</i> , 2018 Chan <i>et al.</i> , 2019 Bowling <i>et al.</i> , 2020
	2010s-	I Tol2 transposase with barcoded GFP Accrual of Tol2-mediated GFP-barcode Insertions Lineage Tree Reconstruction	Single-cell	1000s - 10,000s of cells	None beyond usual scRNA-seq transgene dropout	Transposon mRNA Barcode Accrual Wagner <i>et al.</i> , 2019

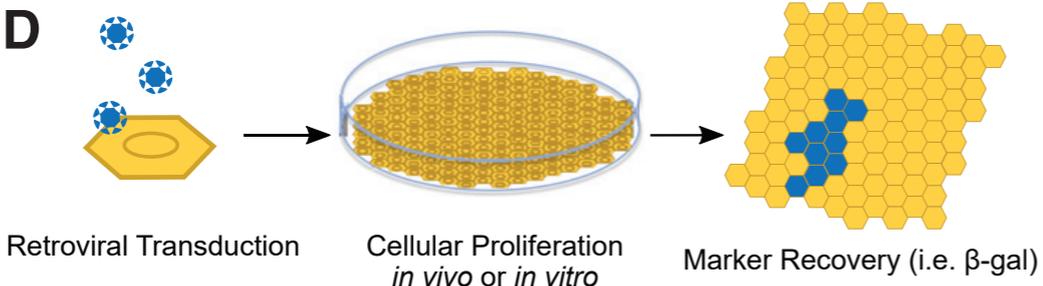
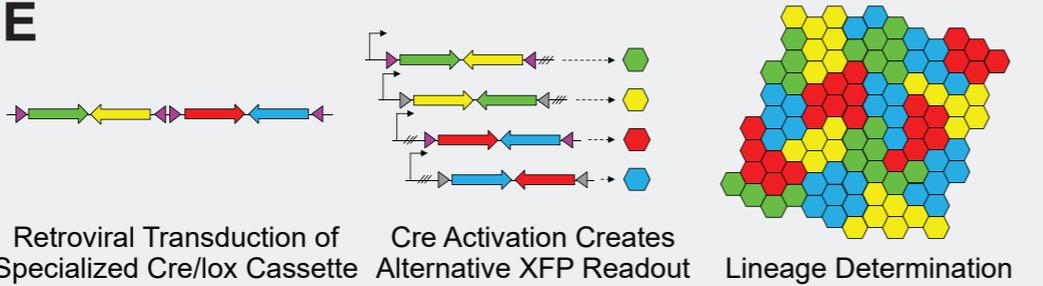
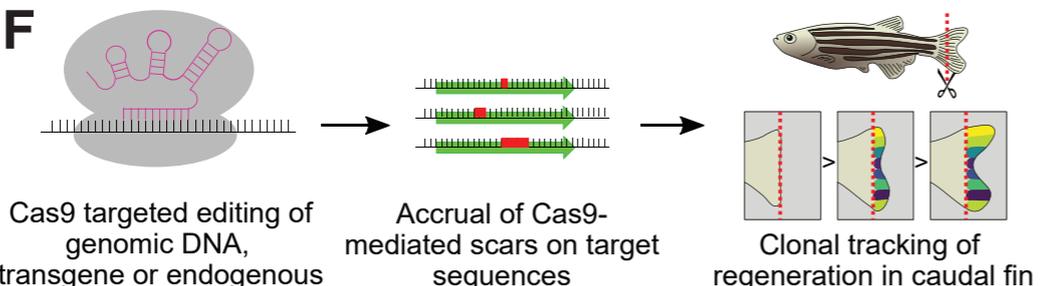


Fate mapping and lineage tracing

Era	Year	Lineage Tracing Technique	Resolution	Scalability	Limitation	Technique & Citation
Observational Biology	1890s	A  <p>Dye Injection 4-cell Stage</p> <p>Observation 8-cell Stage</p> <p>Observation 16-cell Stage</p> <p>Fate Map</p>	Single-cell limited by injection	10s of cells limited by observation	Observational data	Dye Injection and Time Lapse Conklin, 1905 Vogt, 1924
	1920-30s	B  <p>Organizer Engraftment</p> <p>Donor Embryo</p> <p>Recipient Embryo</p> <p>Generation of Secondary Axis</p>	N/A	Tissues	Observational data	Organizer Grafts Spemann and Mangold, 1924 Wetzel, 1929
	1960s	C  <p>8-cell Embryos</p> <p>Chimeric 16-cell Embryo</p> <p>Chimeric Mouse</p>	N/A	Tissues	Only specific to embryo of origin	Chimera Generation Tarkowski, 1965 Mintz, 1965



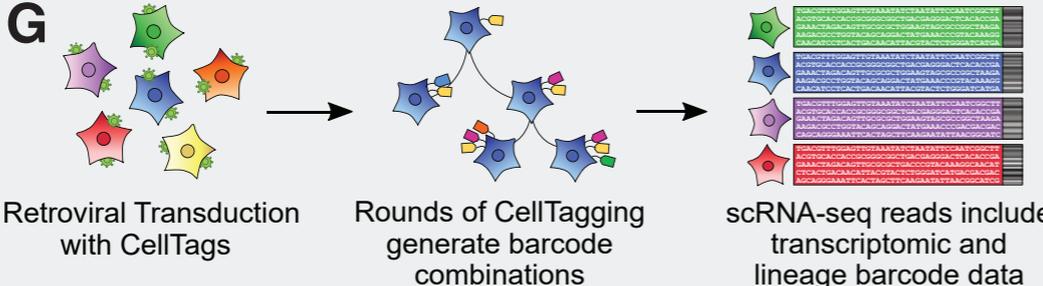
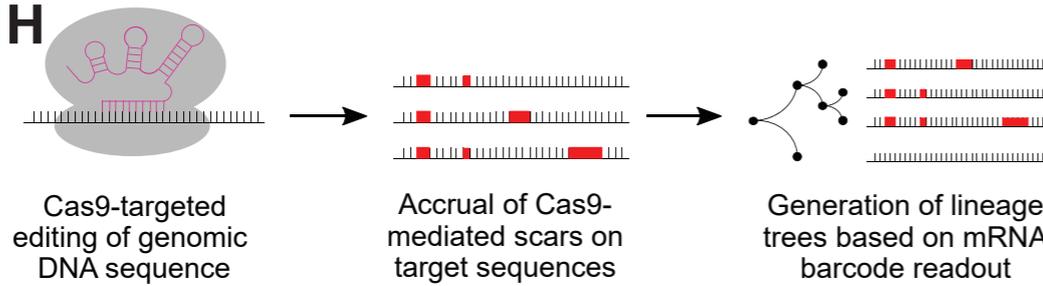
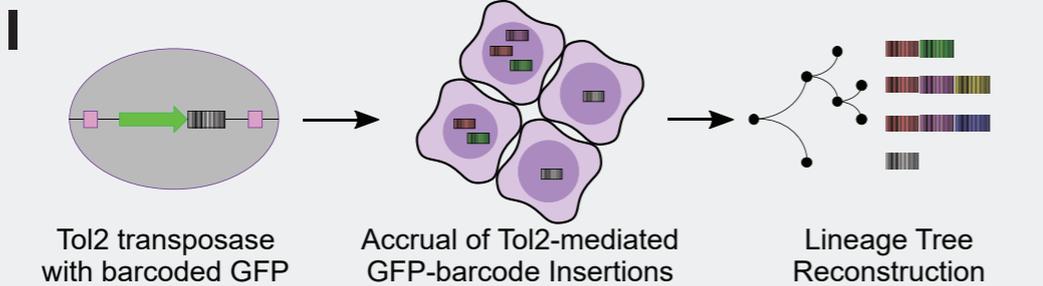
Fate mapping and lineage tracing

Era	Year	Lineage Tracing Technique	Resolution	Scalability	Limitation	Technique & Citation
Molecular Biology	1980s	 <p>D</p> <p>Retroviral Transduction → Cellular Proliferation <i>in vivo</i> or <i>in vitro</i> → Marker Recovery (i.e. β-gal)</p>	Theoretically single clones	10s of cells limited by observation	Observational data	Retroviral Labelling Cepko <i>et al</i> , 1987
	2000s	 <p>E</p> <p>Retroviral Transduction of Specialized Cre/lox Cassette → Cre Activation Creates Alternative XFP Readout → Lineage Determination</p>	Theoretically single clones	100s of cells limited by observation	Observational data	Randomized Recombination Cassettes Livet <i>et al</i> , 2007 Snippert <i>et al</i> , 2010
	2010s	 <p>F</p> <p>Cas9 targeted editing of genomic DNA, transgene or endogenous → Accrual of Cas9-mediated scars on target sequences → Clonal tracking of regeneration in caudal fin</p>	Theoretically single clones	100s of cells limited by observation	Dataset limitation based on collection method	Cas9 Targeted Scar Accrual McKenna <i>et al</i> , 2016 Junker <i>et al</i> , 2017

Still pre-“single-cell revolution” takeover

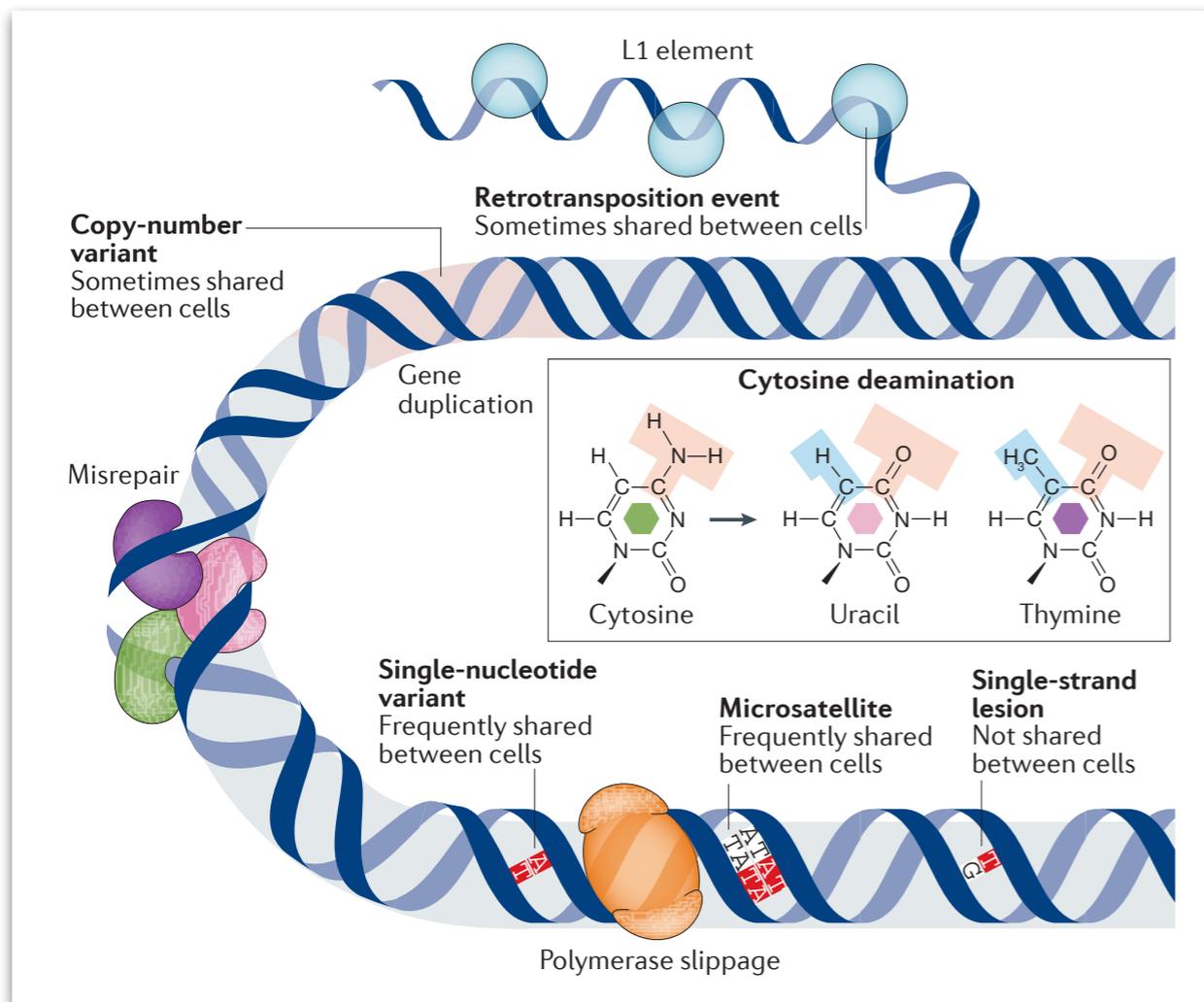


Fate mapping and lineage tracing

Era	Year	Lineage Tracing Technique	Resolution	Scalability	Limitation	Technique & Citation
Single-cell Biology	2010s-	<p>G</p>  <p>Retroviral Transduction with CellTags</p> <p>Rounds of CellTagging generate barcode combinations</p> <p>scRNA-seq reads include transcriptomic and lineage barcode data</p>	Single-cell	1000s - 10,000s of cells	Resolved to clonal and sub-clonal populations	<p>Retroviral mRNA Barcode Accrual</p> <p>Yao <i>et al</i>, 2017</p> <p>Biddy <i>et al</i>, 2018</p> <p>Weinreb <i>et al</i>, 2020</p>
	2010s-	<p>H</p>  <p>Cas9-targeted editing of genomic DNA sequence</p> <p>Accrual of Cas9-mediated scars on target sequences</p> <p>Generation of lineage trees based on mRNA barcode readout</p>	Single-cell	1000s - 10,000s of cells	Information dropout due to Cas9 induced deletion of previous scars	<p>Cas9 mRNA Scars</p> <p>Spanjaard <i>et al</i>, 2018</p> <p>Raj <i>et al</i>, 2018</p> <p>Chan <i>et al</i>, 2019</p> <p>Bowling <i>et al</i>, 2020</p>
	2010s-	<p>I</p>  <p>Tol2 transposase with barcoded GFP</p> <p>Accrual of Tol2-mediated GFP-barcode Insertions</p> <p>Lineage Tree Reconstruction</p>	Single-cell	1000s - 10,000s of cells	None beyond usual scRNA-seq transgene dropout	<p>Transposon mRNA Barcode Accrual</p> <p>Wagner <i>et al</i>, 2019</p>



Using somatic mutations



From Figure 4 of Woodworth et al., Nat Rev Gen (2017)

Advantage: already “in the data” or “free”

Two main **limitations** of using somatic variation in lineage tracing:

1. **WGS** required (\$\$\$, unscalable)
2. Inherent read **sparsity** of scRNA-seq



Using mtDNA mutations

Article **Cell**

Lineage Tracing in Humans Enabled by Mitochondrial Mutations and Single-Cell Genomics

Leif S. Ludwig,^{1,2,15,*} Caleb A. Lareau,^{1,2,3,4,15} Jacob C. Ulirsch,^{1,2,4,15} Elena Christian,¹ Christoph Muus,^{1,5} Lauren H. Li,^{1,2} Karin Pelka,^{1,6,7} Will Ge,¹ Yaara Oren,^{1,8} Alison Brack,¹ Travis Law,¹ Christopher Rodman,¹ Jonathan H. Chen,^{1,9} Genevieve M. Boland,^{6,10} Nir Hacohen,^{1,6,7} Orit Rozenblatt-Rosen,¹ Martin J. Aryee,^{1,3,11} Jason D. Buenrostro,^{1,12} Aviv Regev,^{1,13,*} and Vijay G. Sankaran^{1,2,14,16,*}

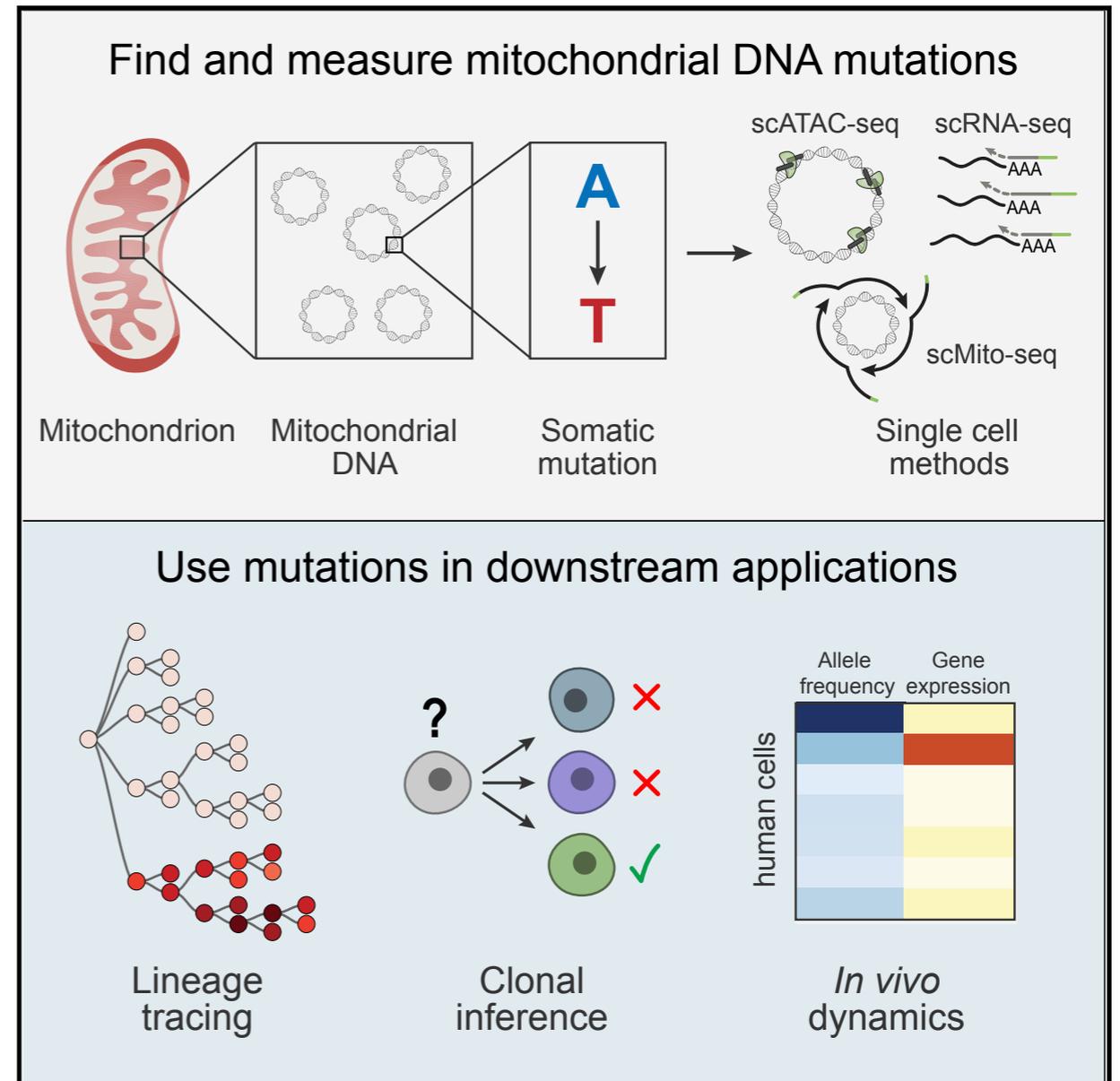
Advantages:

1. Again, “free”
2. mtDNA mutation rates > gDNA mutation rates
3. Readily paired GEX / CA with clonal lineage information
4. No WGS or fancy barcoding required

Limitations:

1. Current approaches offer limited coverage of mito genome
2. Potential for horizontal gene transfer in mito genome (unclear to what extent)

Graphical Abstract



From Graphical Abstract of Ludwig et al., Cell (2021)



GESTALT (genome editing of synthetic target arrays for lineage tracing)

- This paper sets a precedent for large-scale lineage tracing - how? Tracing must...

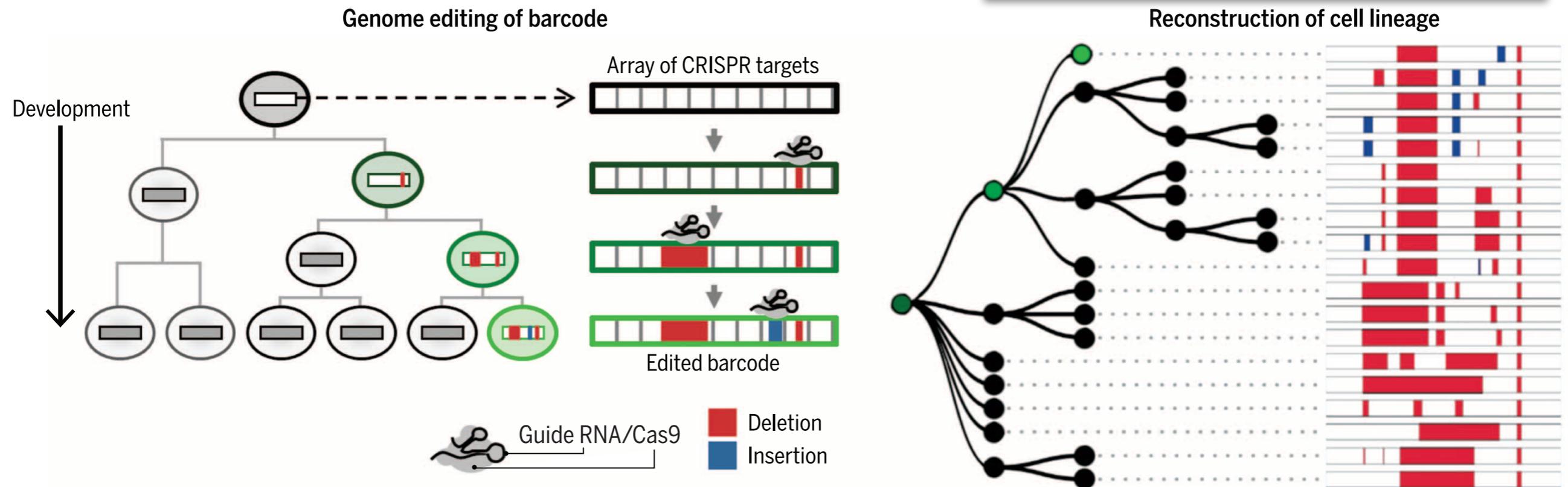
1. Impart unique marks over division
2. Marks must accumulate over time
3. Easy single-cell readout

RESEARCH ARTICLE

CELL LINEAGE TRACING

Whole-organism lineage tracing by combinatorial and cumulative genome editing

Aaron McKenna,^{1*} Gregory M. Findlay,^{1*} James A. Gagnon,^{2*} Marshall S. Horwitz,^{1,3} Alexander F. Schier,^{2,4,5,6†} Jay Shendure^{1,7†}

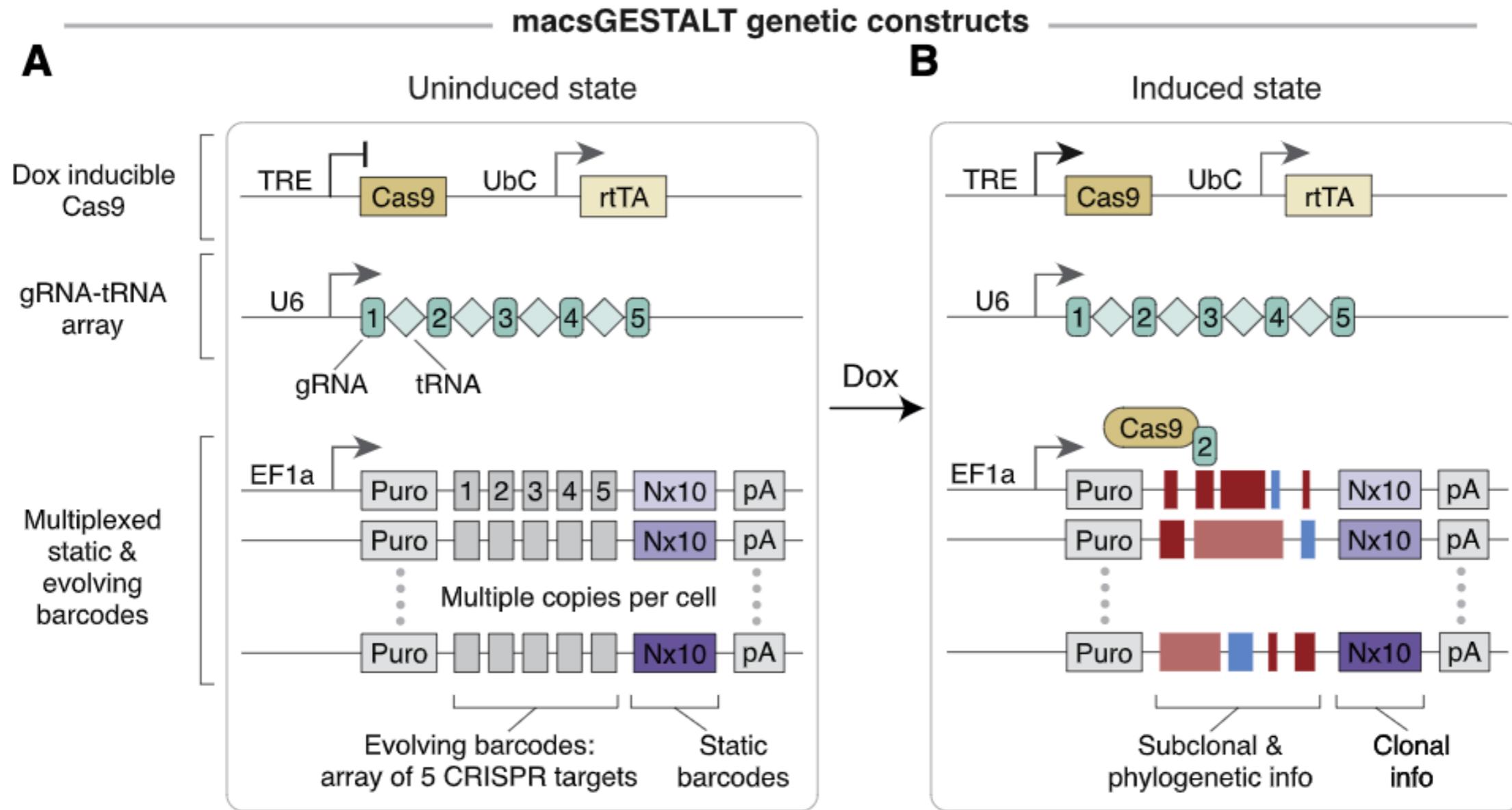


- Applied to cell culture and zebrafish
- Found that most cells in adult organs derive from relatively few embryonic ancestor cells

From Summary Figure of McKenna et al., Science (2016)



macsGESTALT for high-res lineage tracing



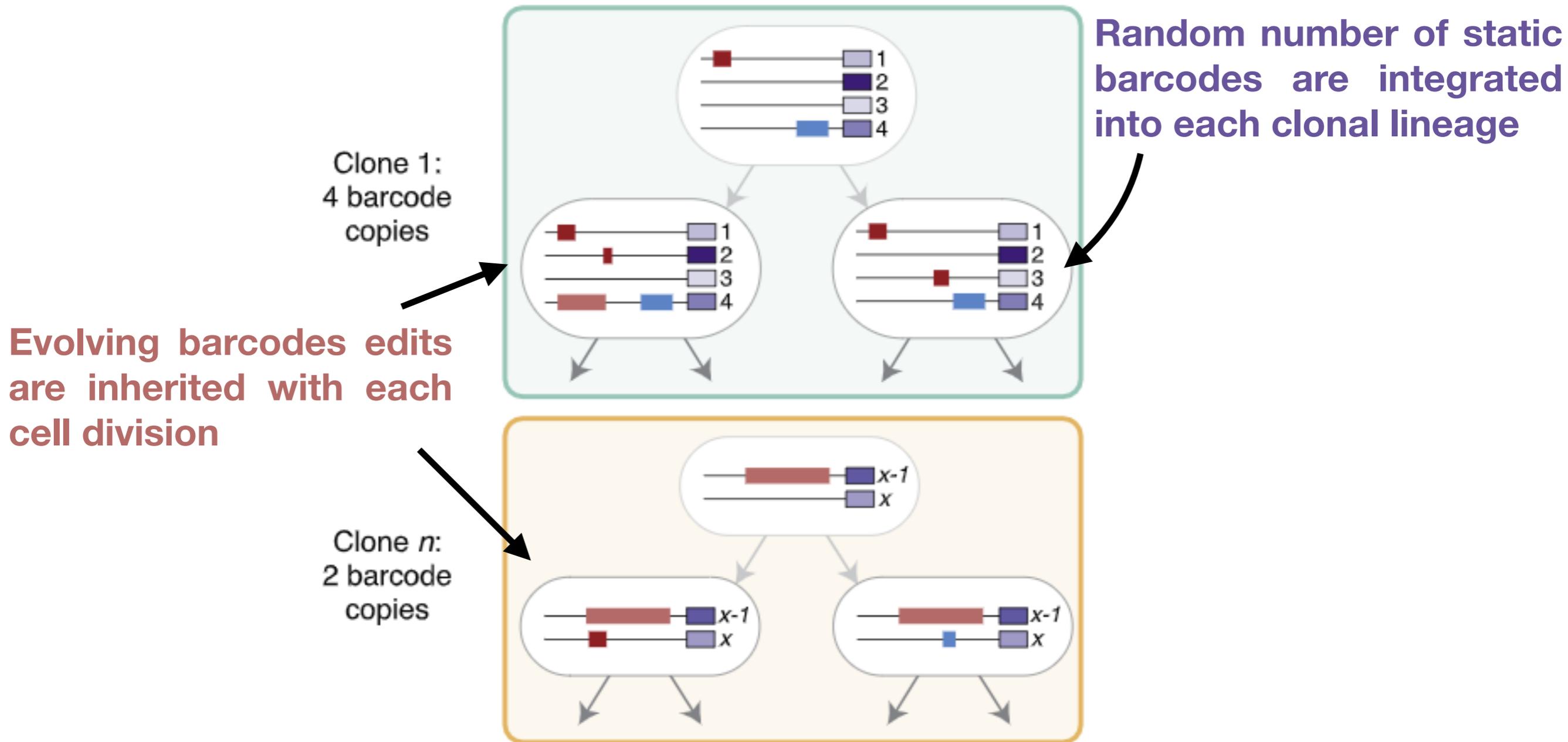
Evolving barcodes (via indel mutagenesis) track sub-clonal phylogenetic information



macsGESTALT for high-res lineage tracing

Behavior in cells

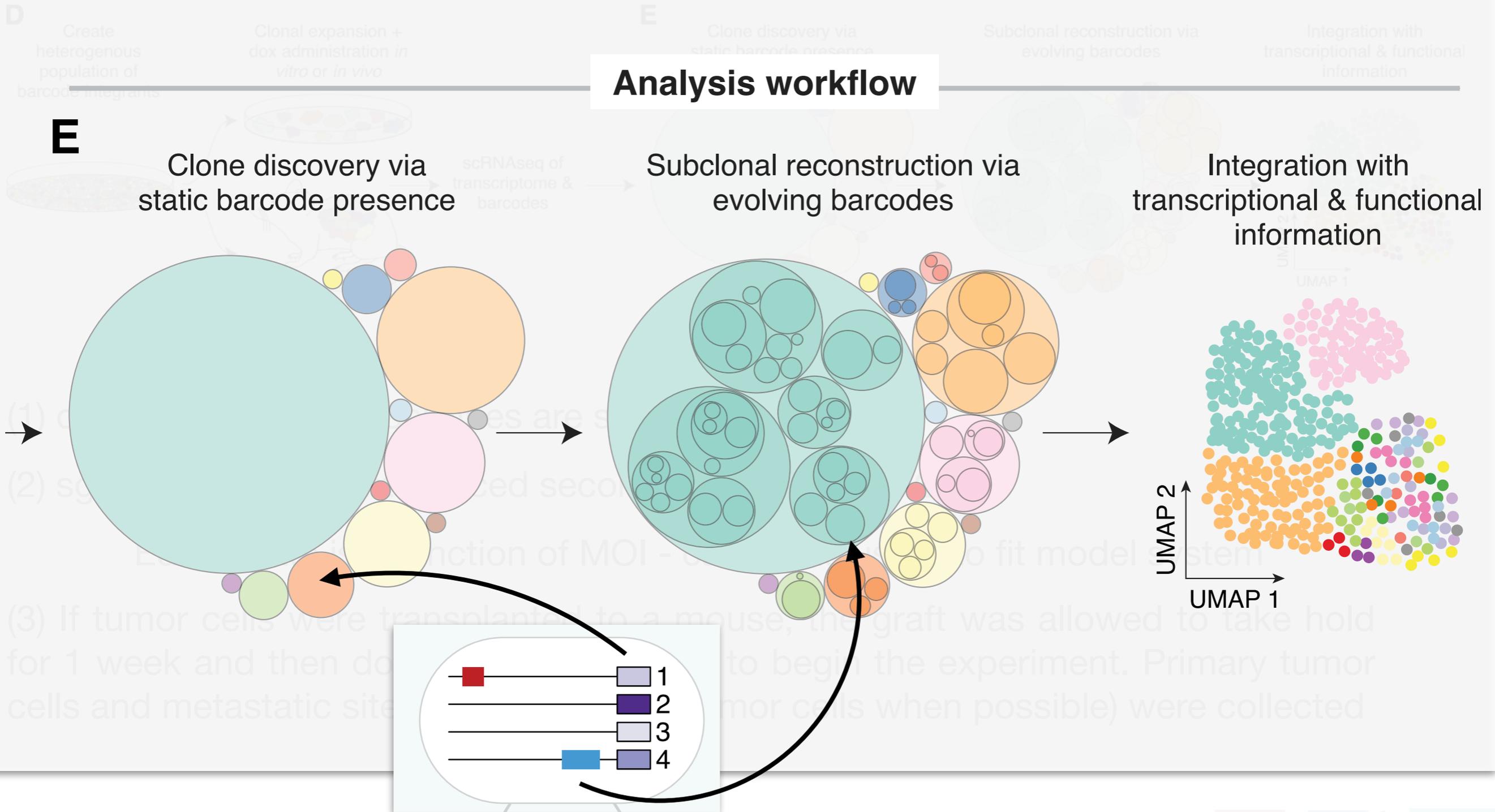
C Population of n clones and x barcodes



macsGESTALT for high-res lineage tracing

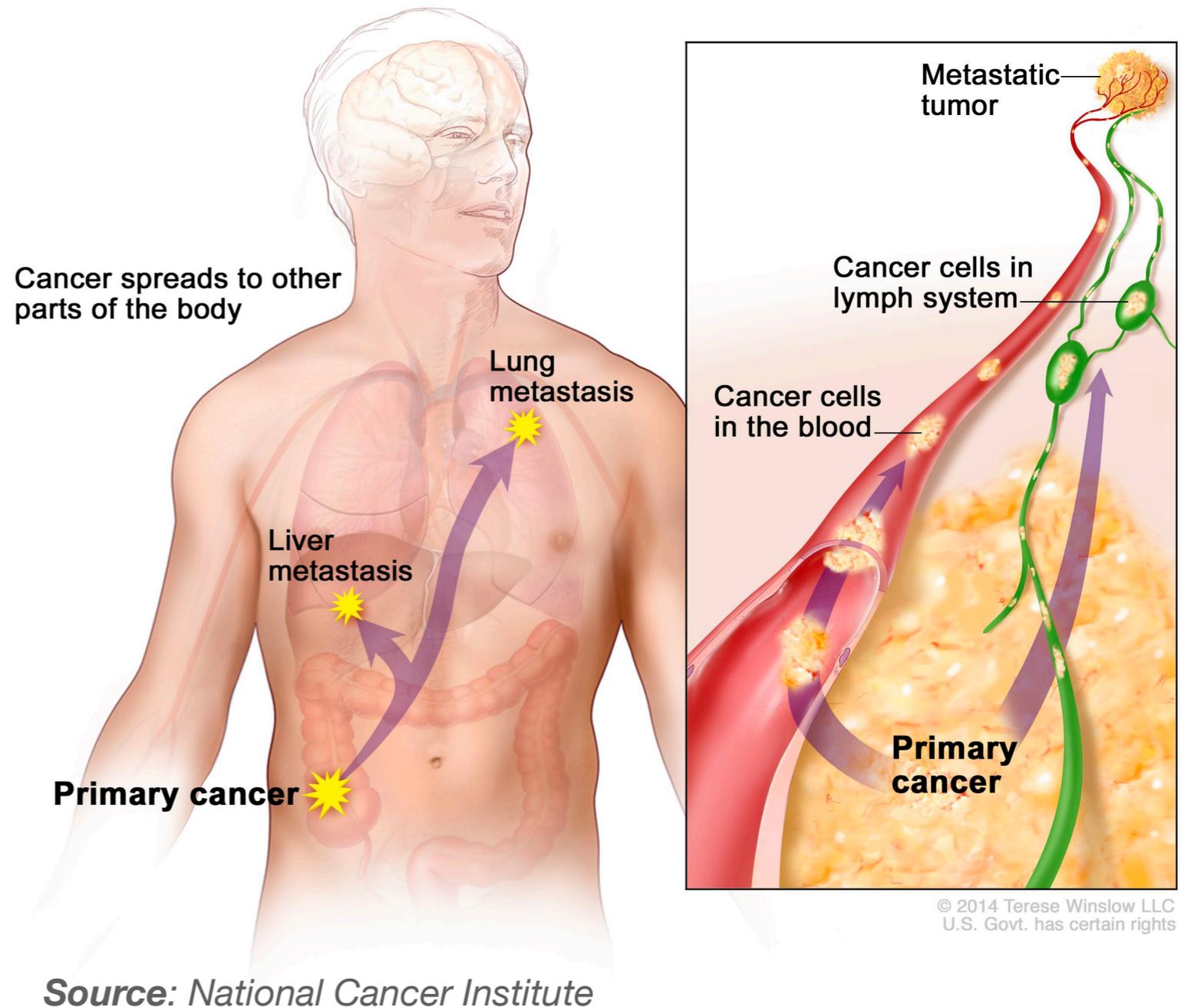
Experimental workflow

Analysis workflow



What is metastasis and EMT?

- Metastasis = most cancer deaths
- EMT ~ metastasis
- EMT thought to play a role across many cancer types



Most metastases arise from rare clones

Experiment: combine macsGESTALT with scRNA-seq



From Figure 2 of Simeonov et al., Cancer Cell (2021)

PDAC is very deadly (5-year survival rate of 9%)

KPCY mouse tumor model cell line transplanted into non-tumor-bearing mice

Good model for two reasons:

- (1) This model exhibits consistent metastasis kinetics
- (2) Good model of human disease ($Kras$ GoF and $p53$ LoF are the most common drivers of human PDAC)
- (3) minimal *in vitro* cell line culture time
- (4) pancreatic focal lesion disseminates to the same sites as in human PDAC (incl. liver and lung)

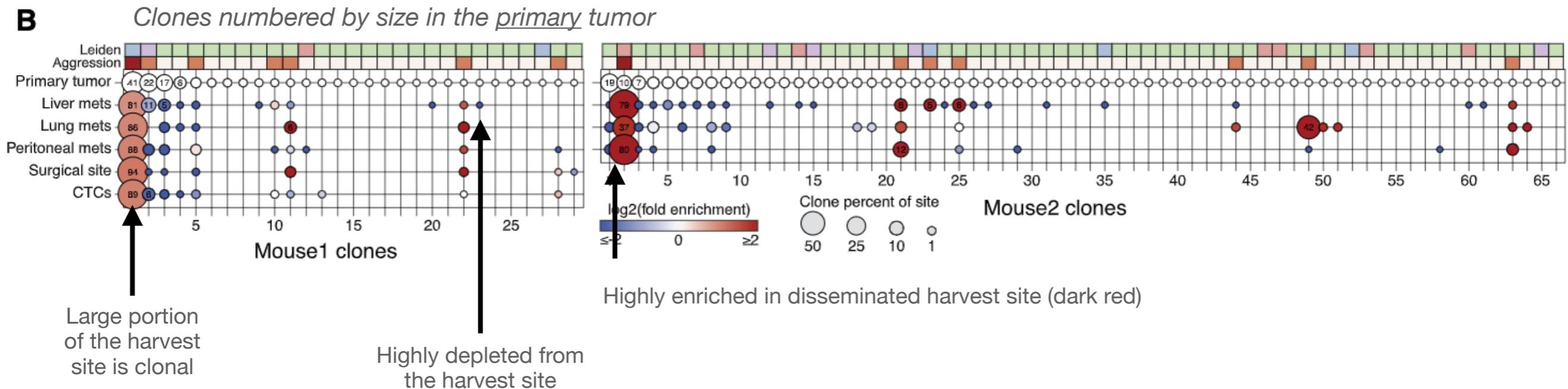


Most metastases arise from rare clones

Clonal reconstruction via static (purple) barcodes:

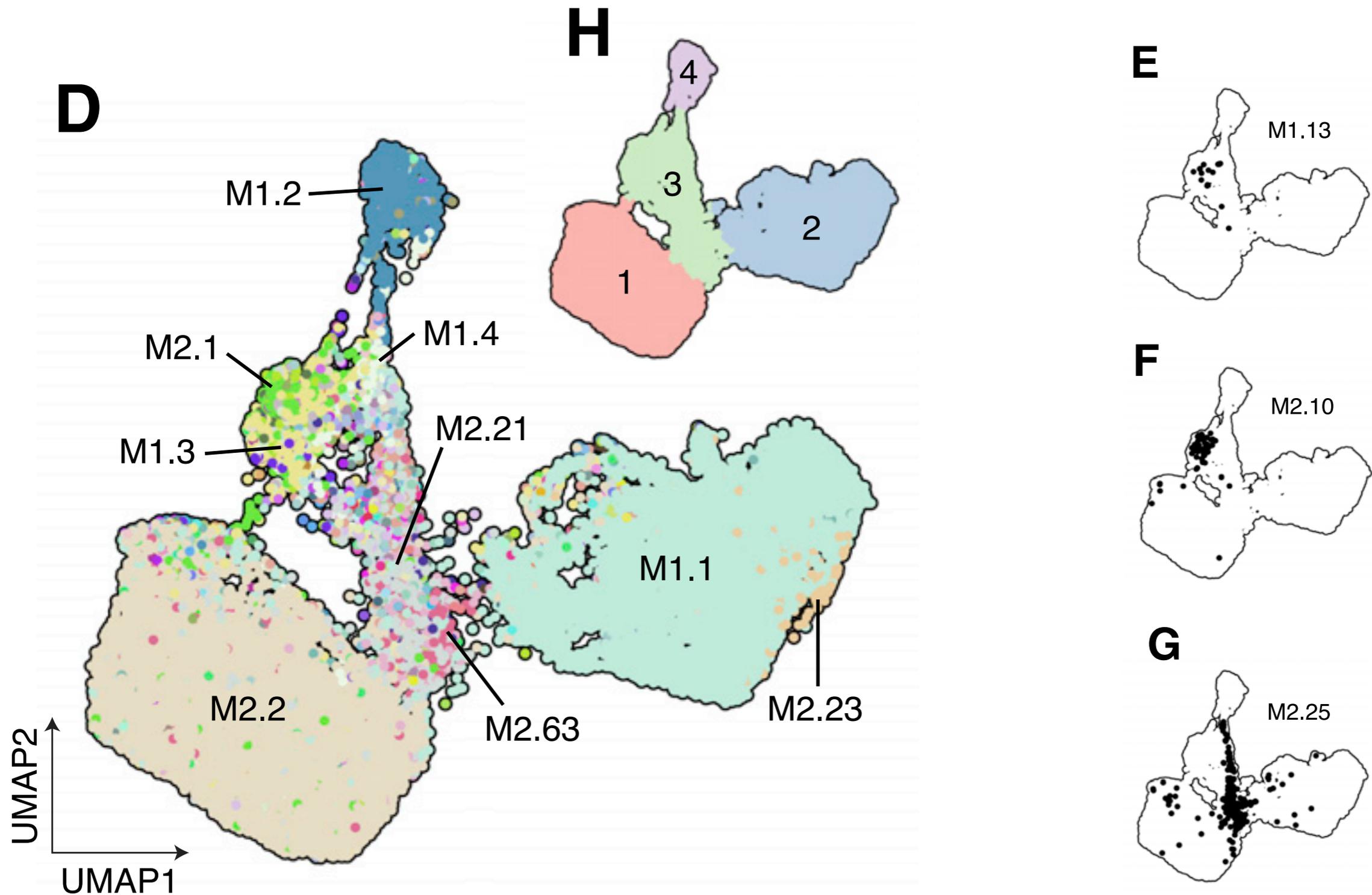
Circle size = % contribution to harvest site

Circle color = enrichment compared to primary tumor

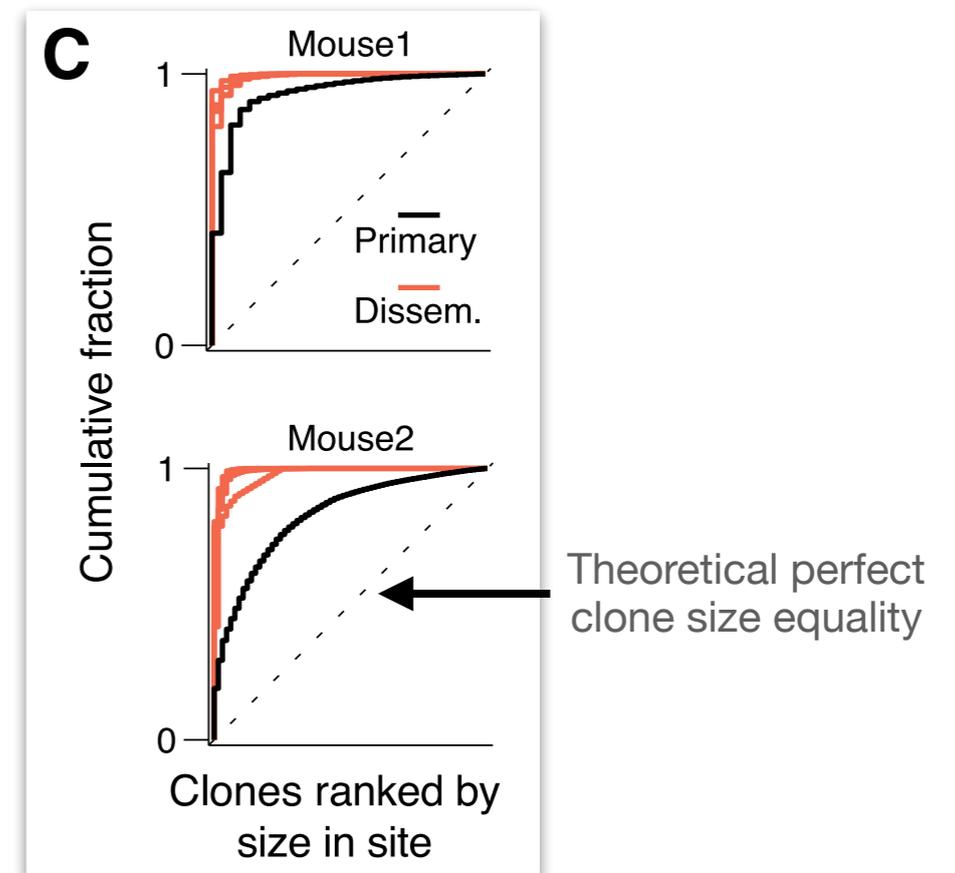
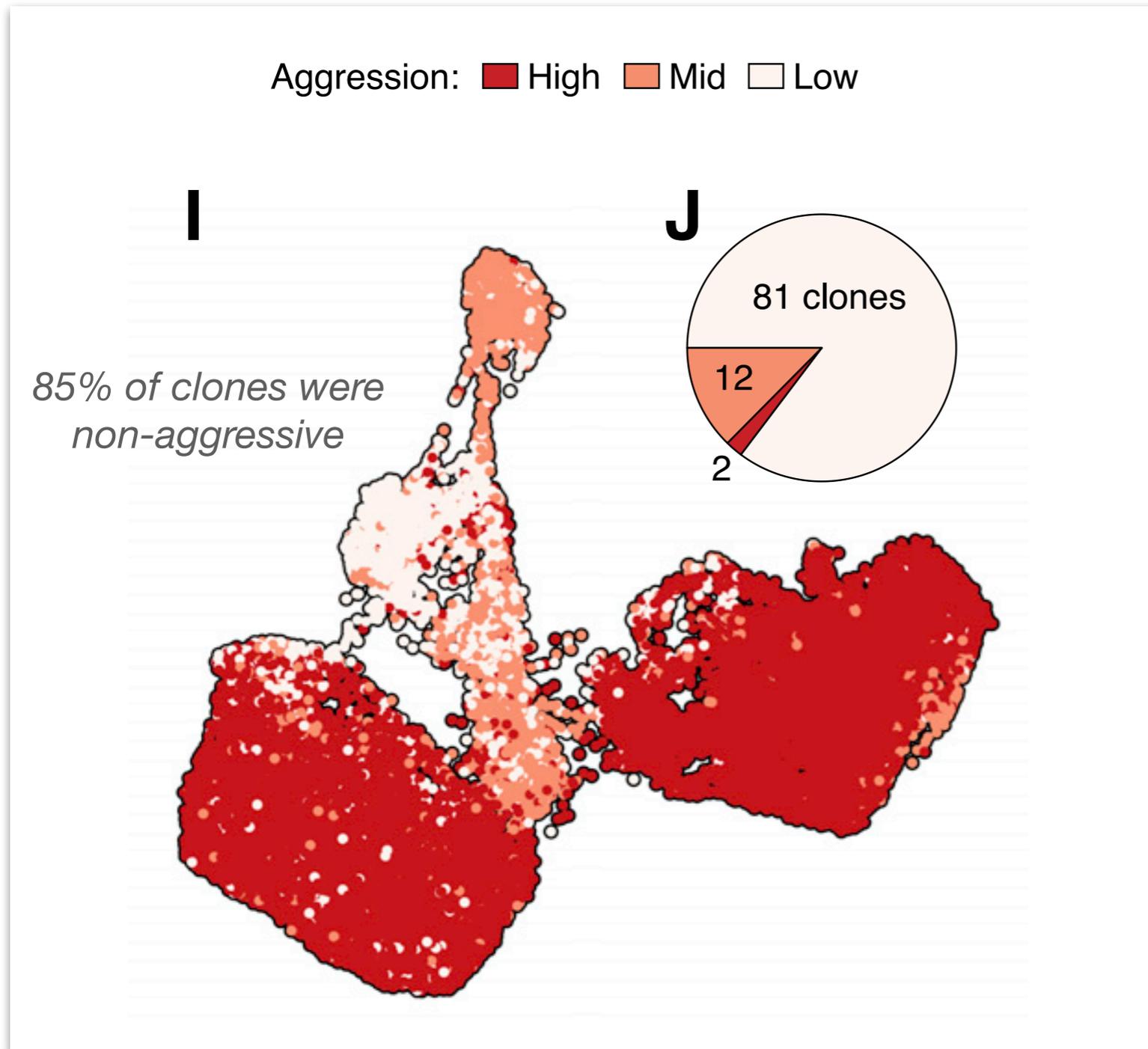


“51% of clones (48/95) failed to metastasize at all, suggesting that mutations in *Kras* and *p53* alone do not ensure metastatic success.”

Most metastases arise from rare clones

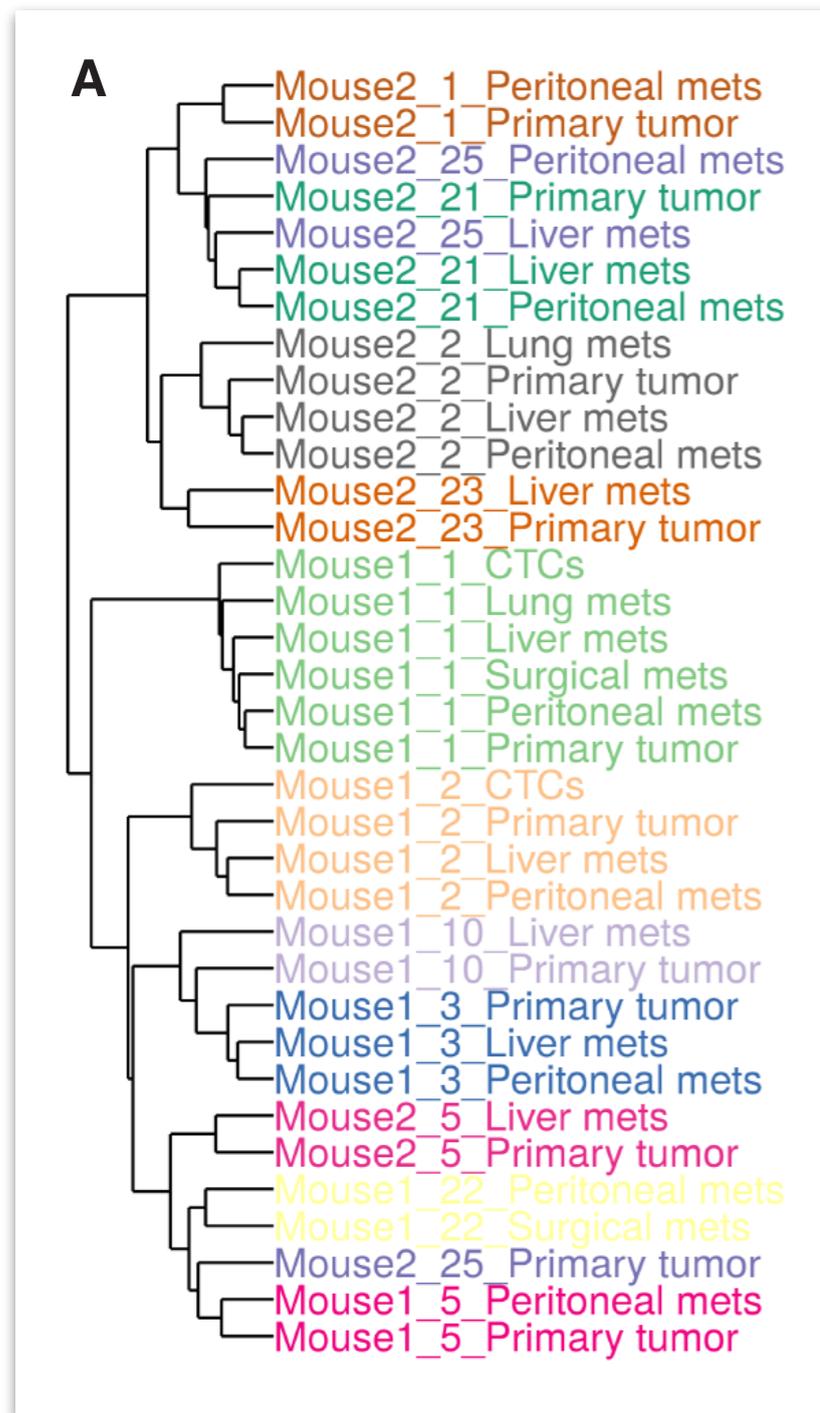


Most metastases arise from rare clones

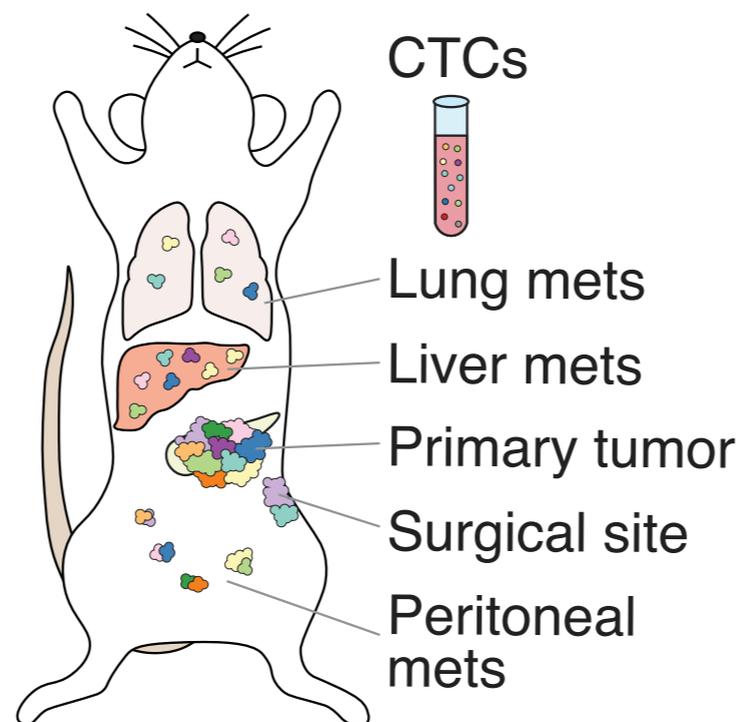


Just **two clones** dominate most of the cell population!

Cells retain transcriptional identity after metastasis



- Clone-site pseudobulk analysis - samples colored by clone
- Samples hierarchically clustered (based on scRNA-seq) - cluster preferentially by clone rather than harvest site

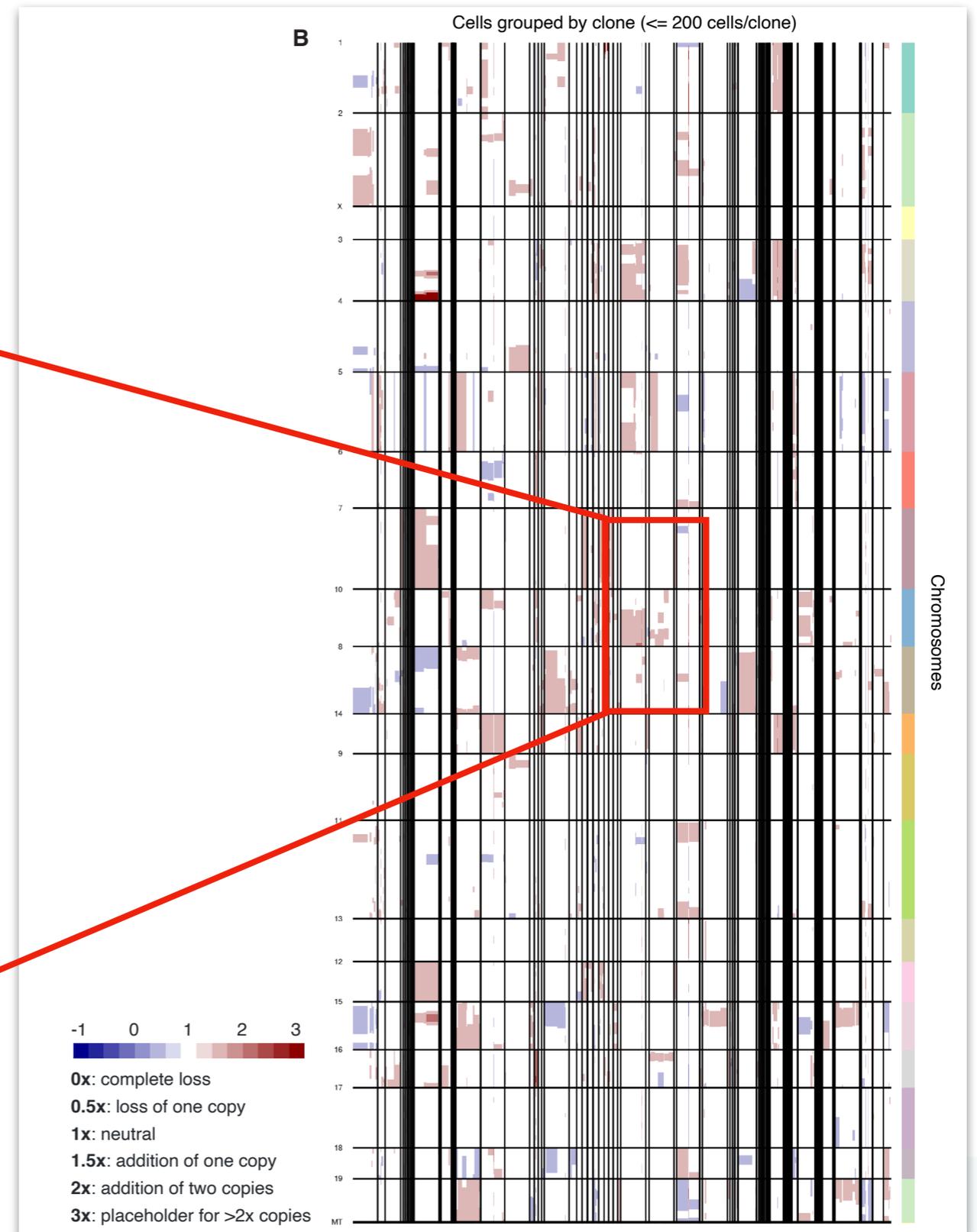
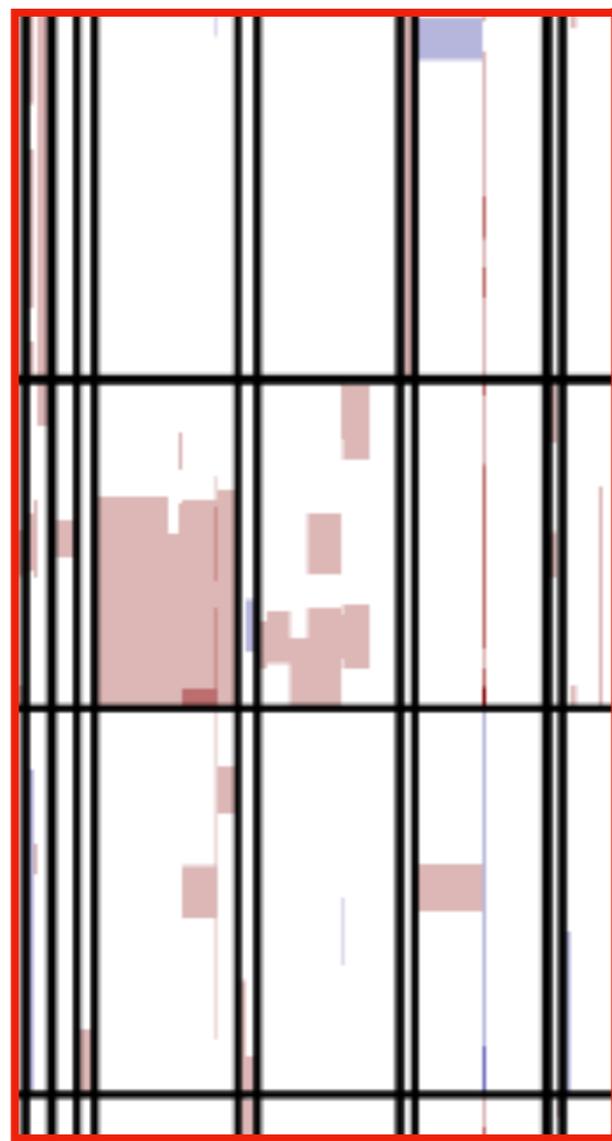


- ① FACS sort
GFP+
PDAC cells
- ② sc-Seq
transcriptome
& barcodes

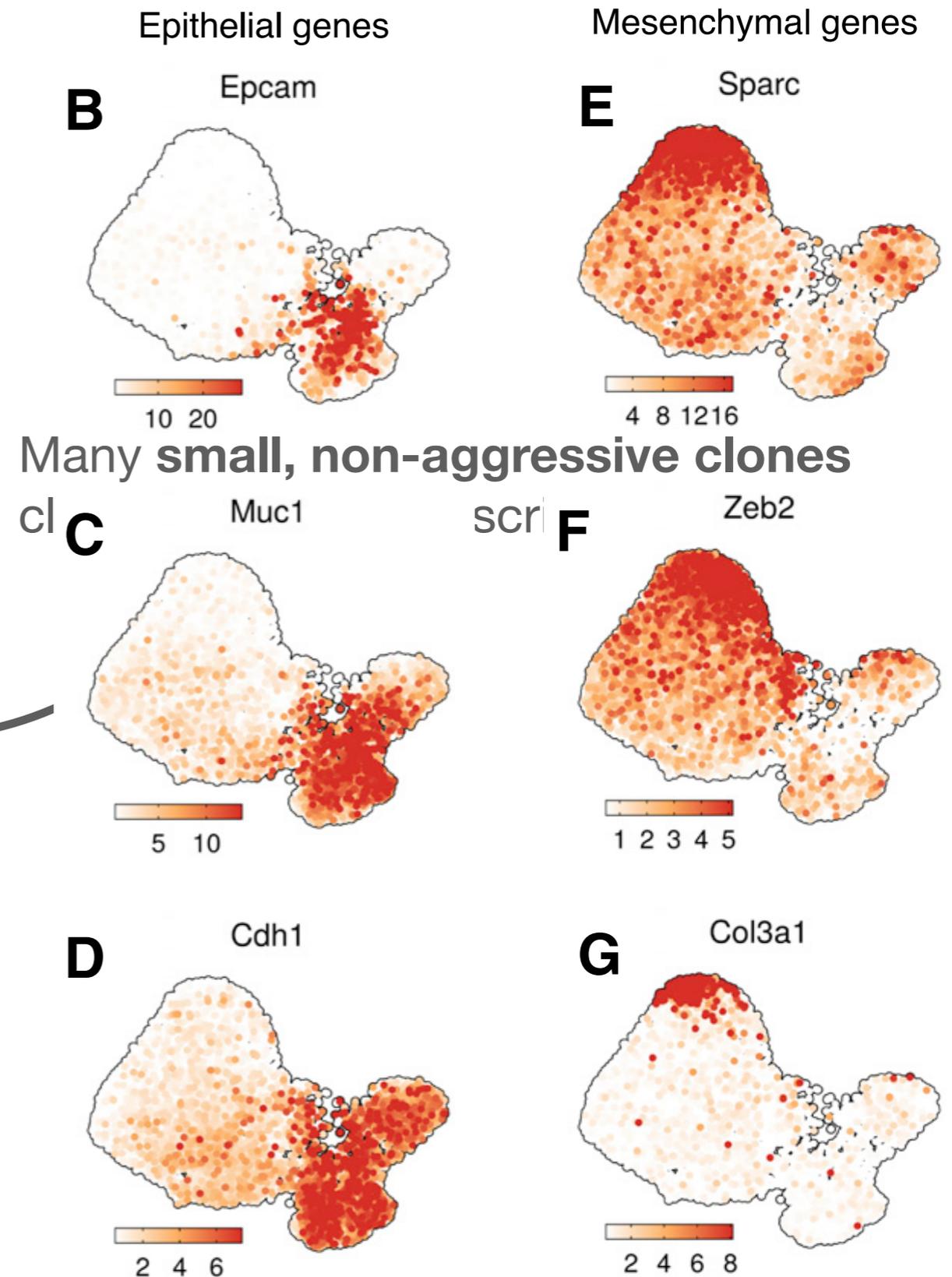
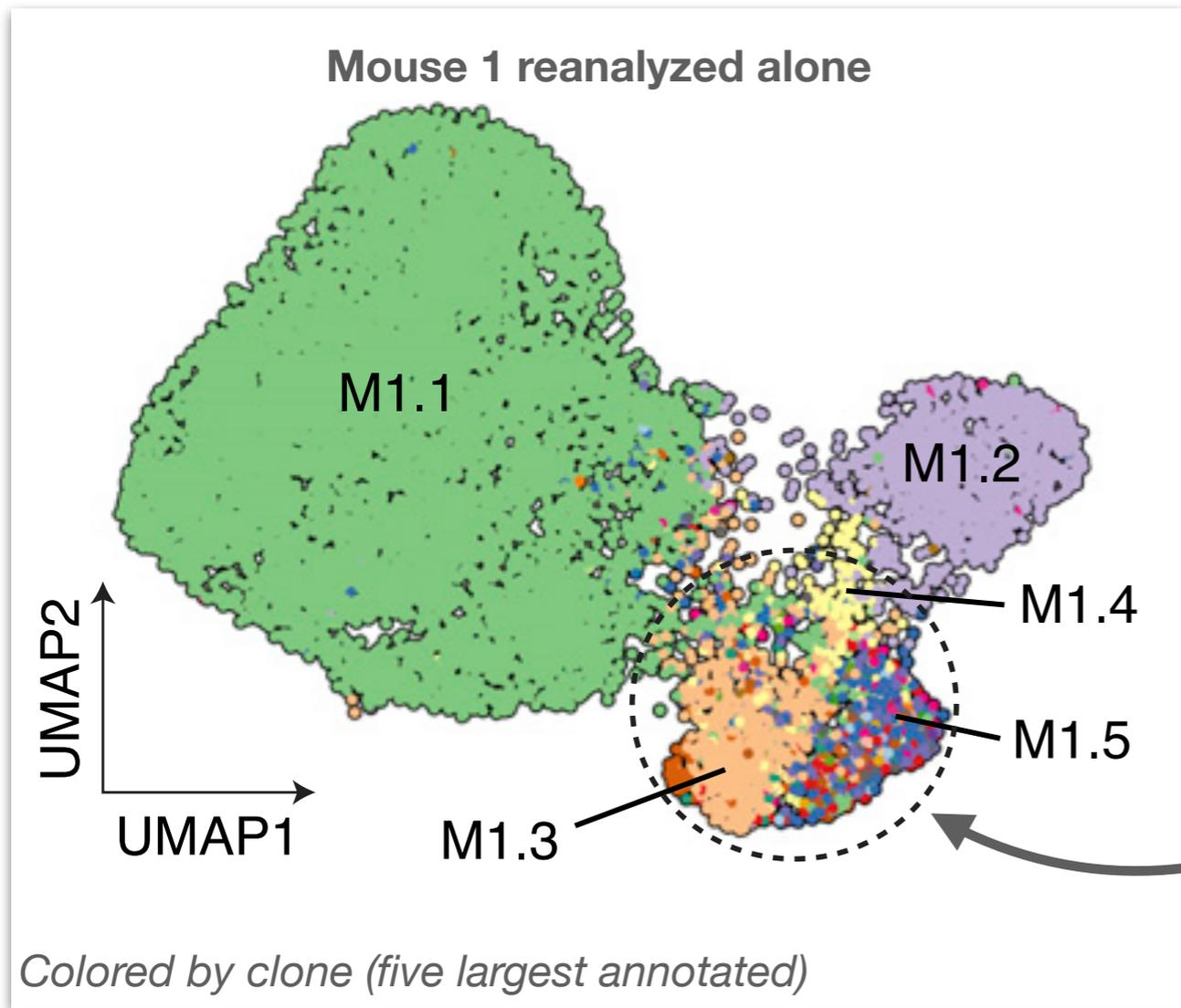


Cells retain transcriptional identity after metastasis

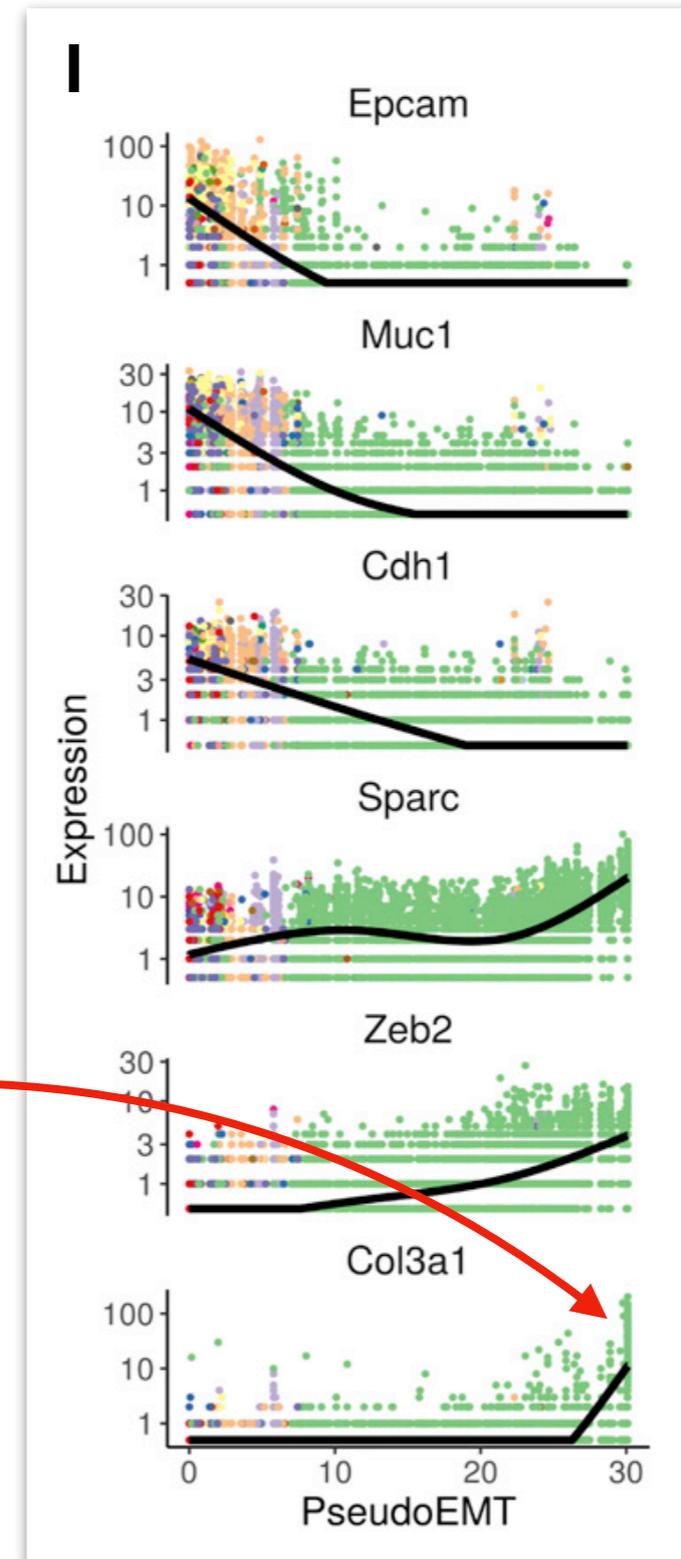
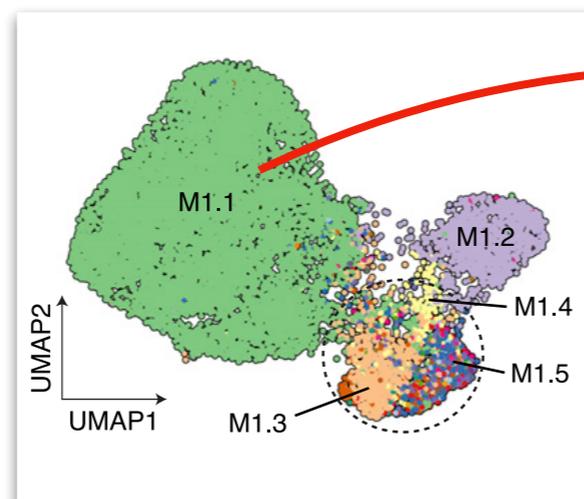
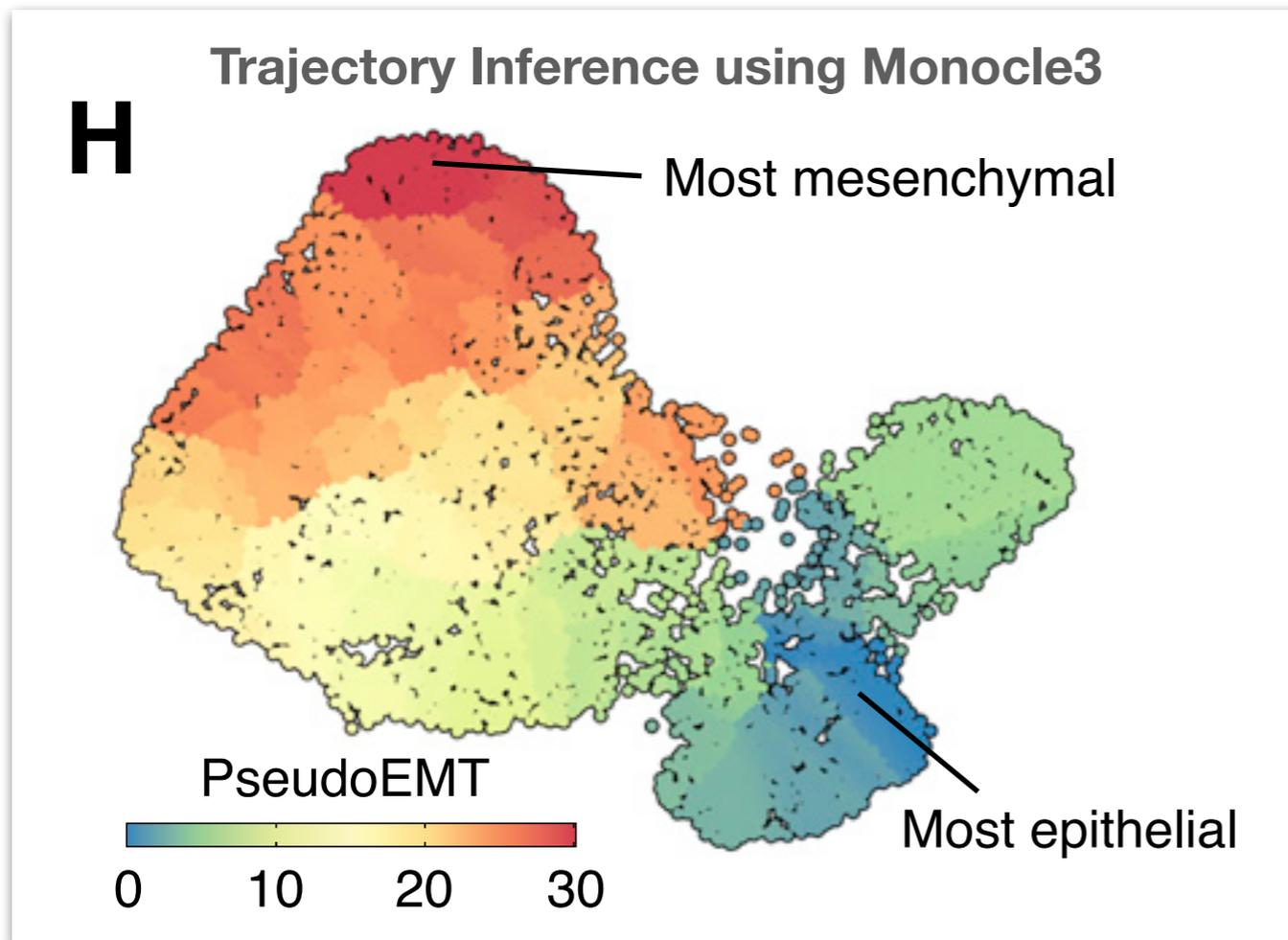
- Performed *InferCNV* analysis



Transcriptional EMT continuum *in vivo*

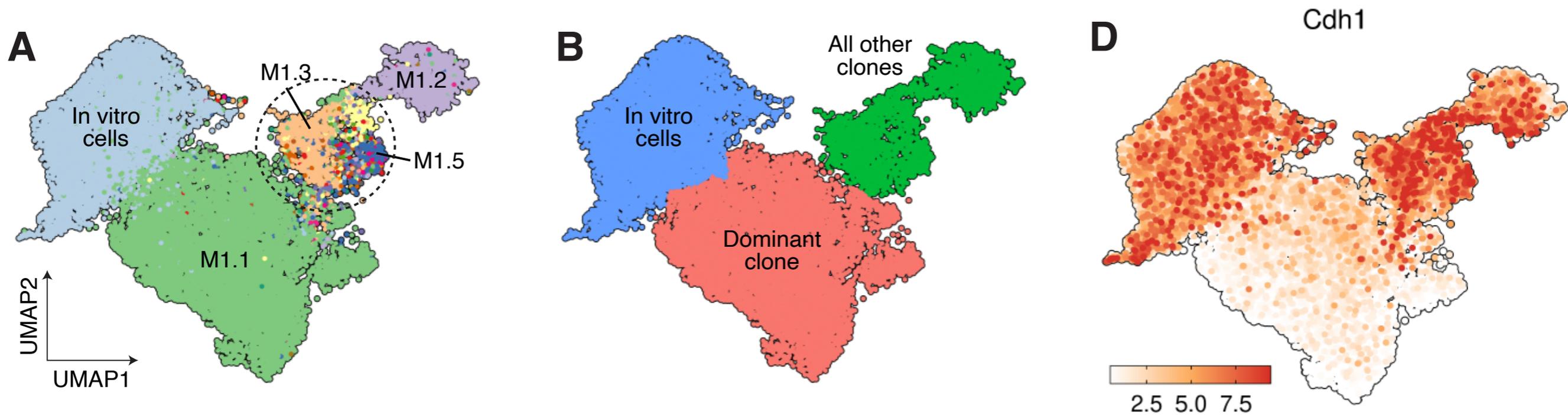


Transcriptional EMT continuum *in vivo*

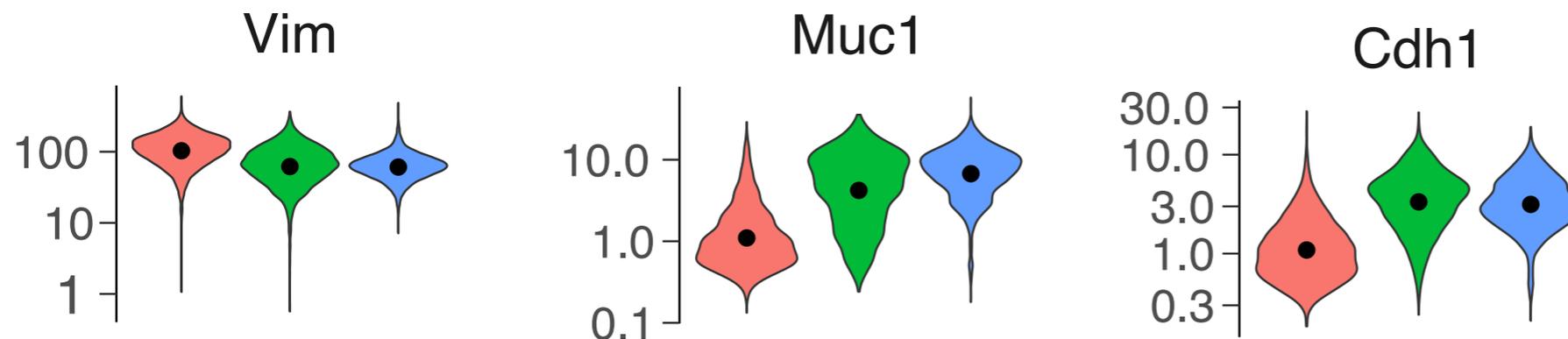


Transcriptional EMT continuum *in vivo*

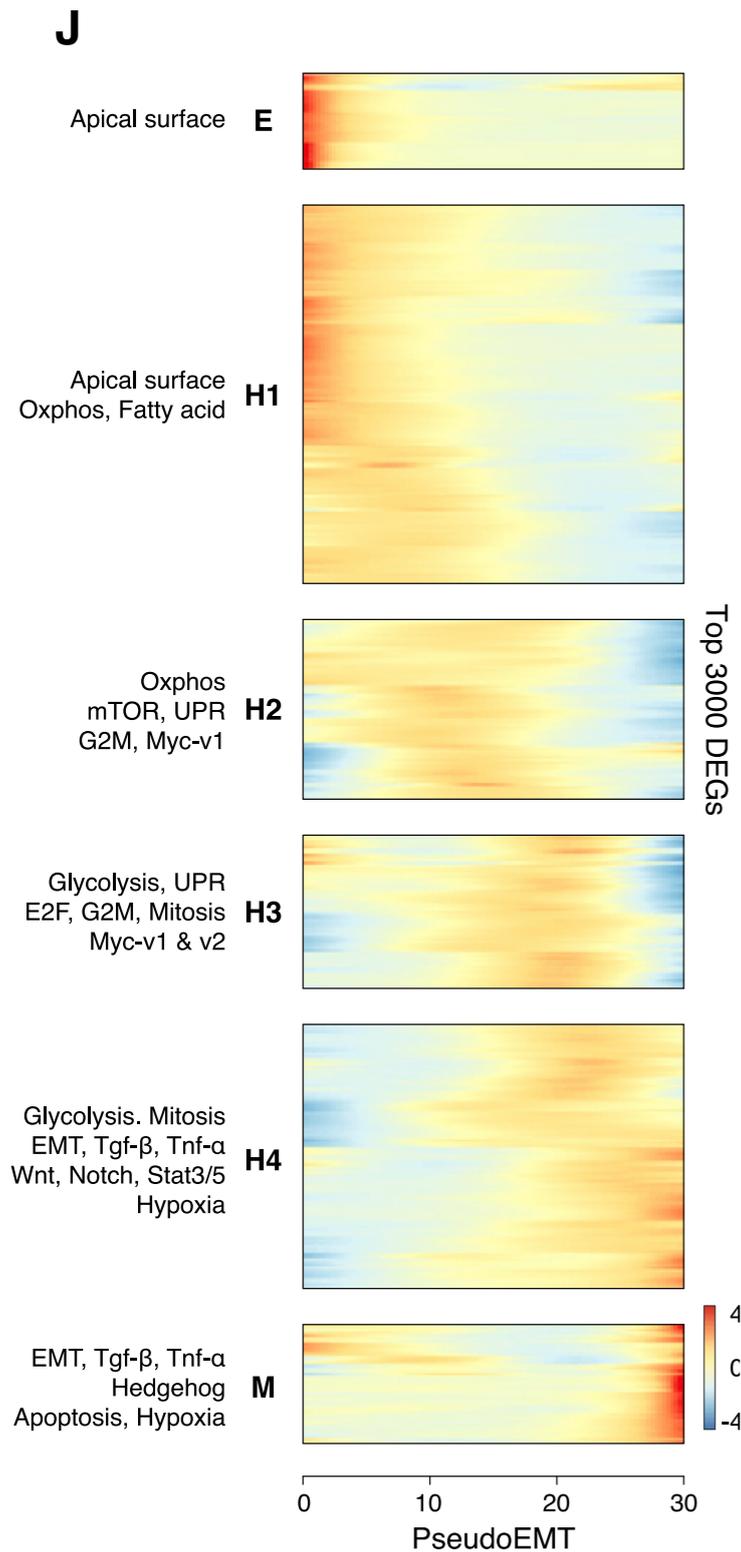
- 27/29 clones are epithelial - is this the default transcriptional state?
- scRNA-seq of *in vitro* cultured cells -> 40 distinct clones



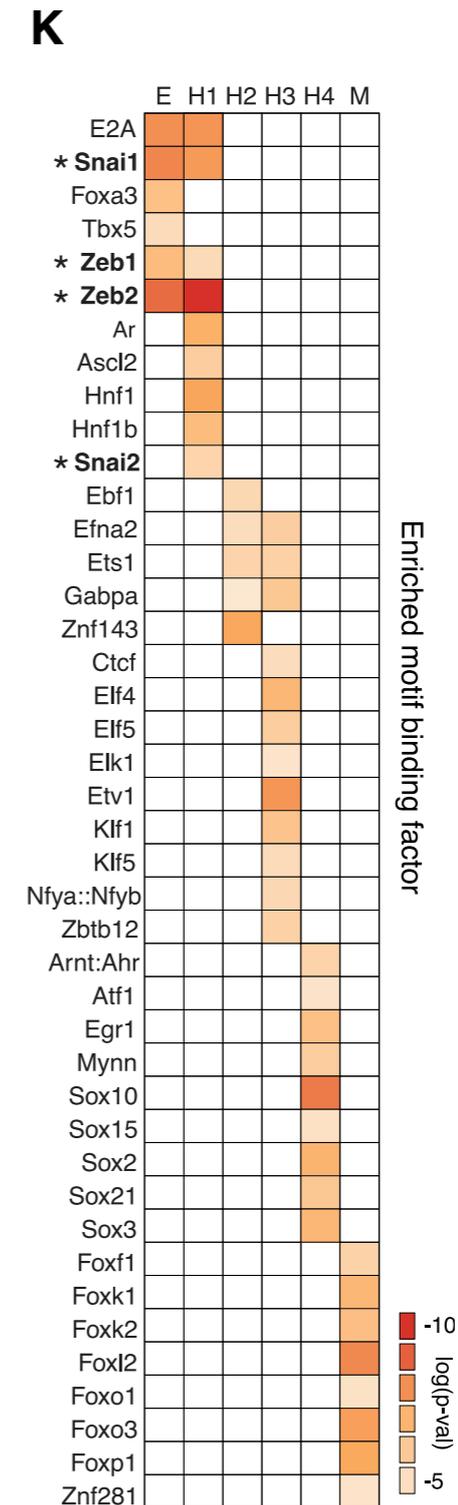
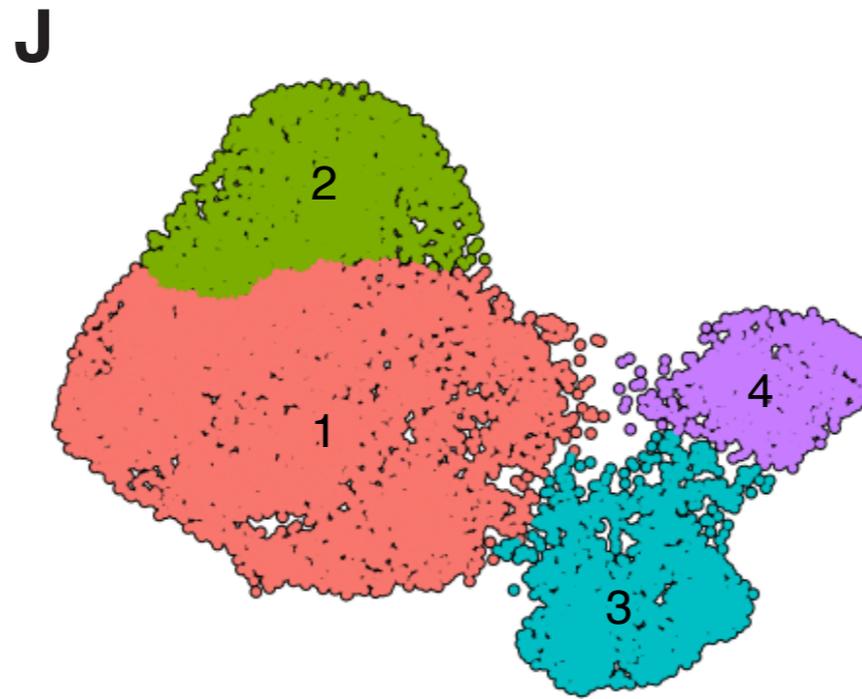
- Compared to PDAC / EMT, *in vitro* cultured cells **strikingly epithelial**



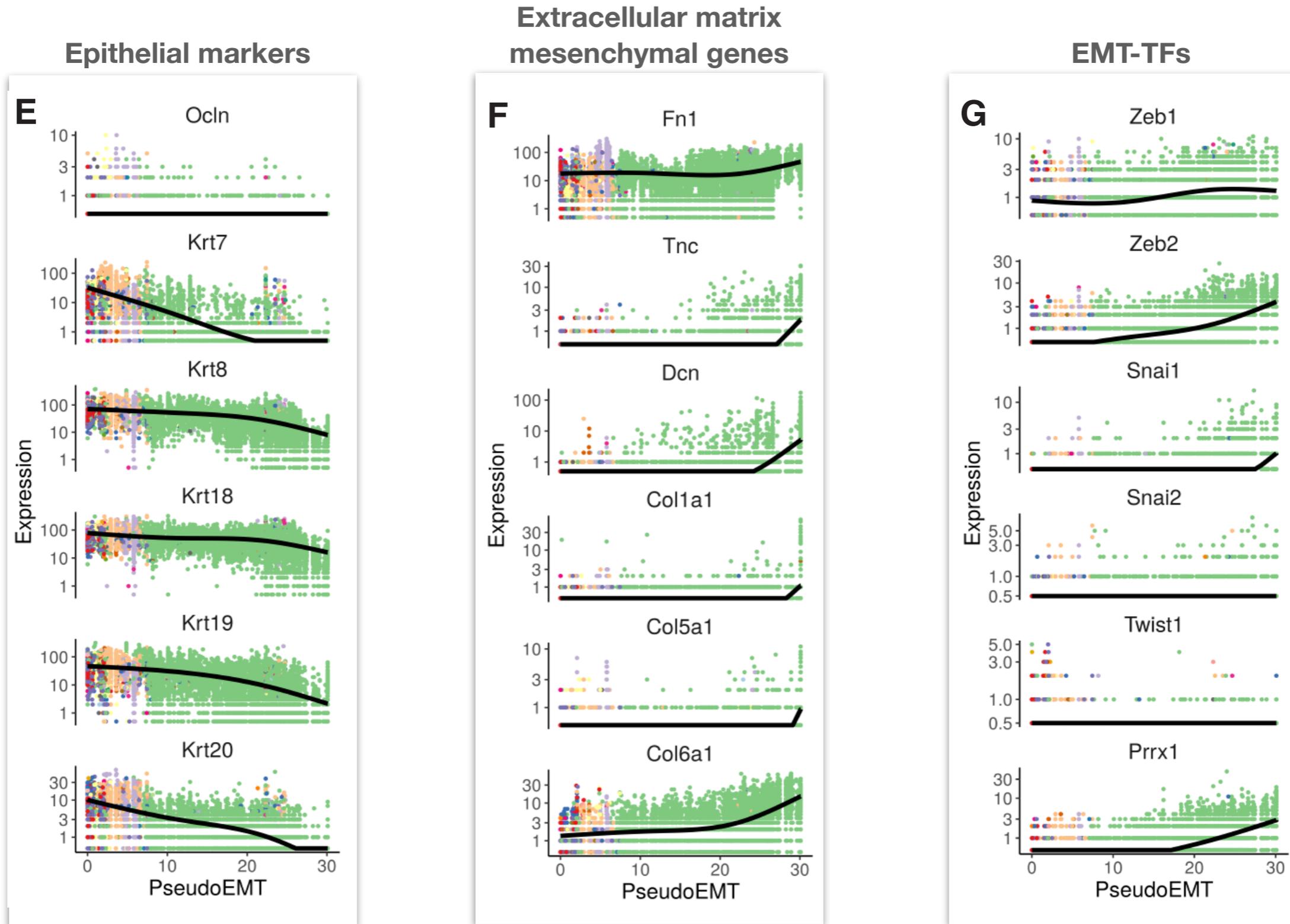
Transcriptional EMT continuum *in vivo*



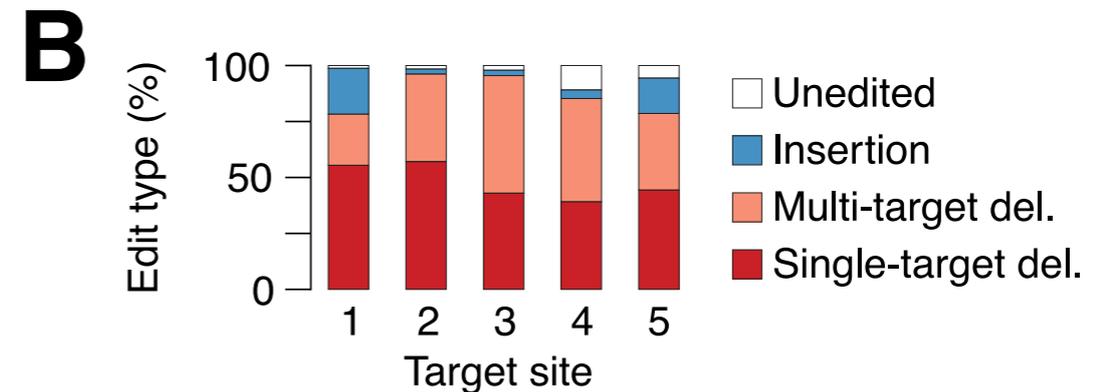
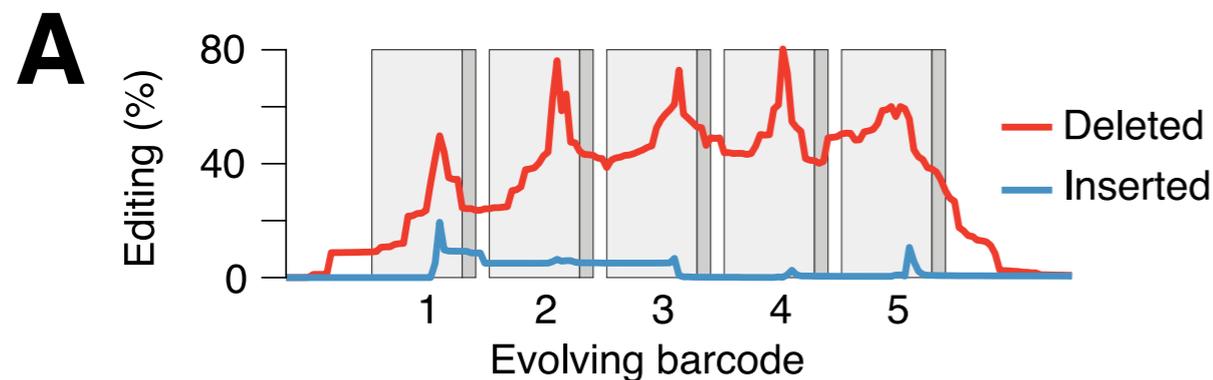
- Remember: not an EMT model but a PDAC model
- Cancer proliferation ~ EMT
- Possible ties between common cancer targets like Myc / mTOR / Wnt / Notch / etc and EMT



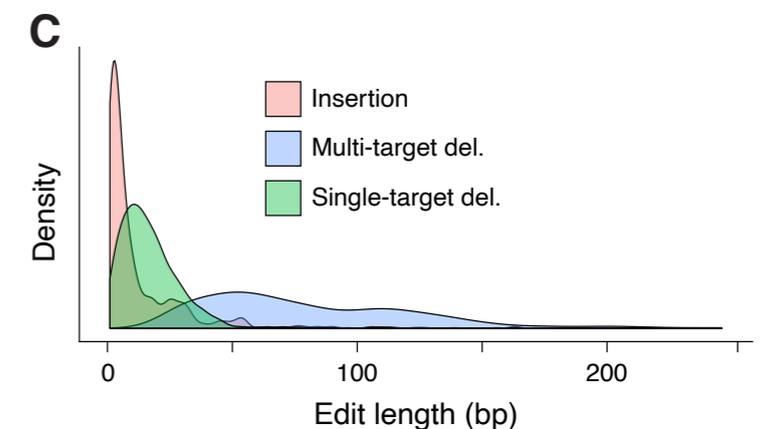
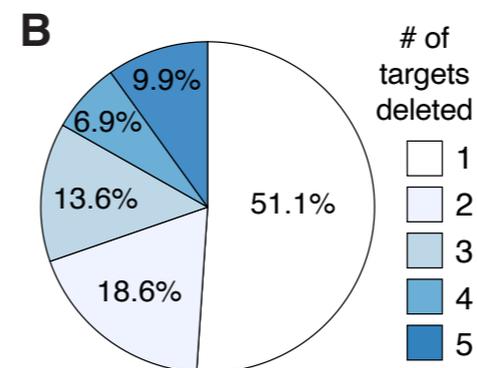
Transcriptional EMT continuum *in vivo*



High-res subclonal lineage reconstruction



	Mouse 1	Mouse 2	Both mice
Total barcodes	50,269	26,705	76,974
Total barcodes with edit	50,132	25,968	76,100
% of barcodes with edit	99.7%	97.2%	98.9%
Total targets	251,345	133,525	384,870
Total targets with edit	246,232	122,013	368,245
% of targets with edit	98.0%	91.4%	95.7%
Distinct edits	2,104	1,383	3,487 (633)
Distinct evolving barcodes	3,886	1,696	5,582 (61)
Distinct barcode-of-barcodes (subclones)	3,997	2,062	6,059 (4)

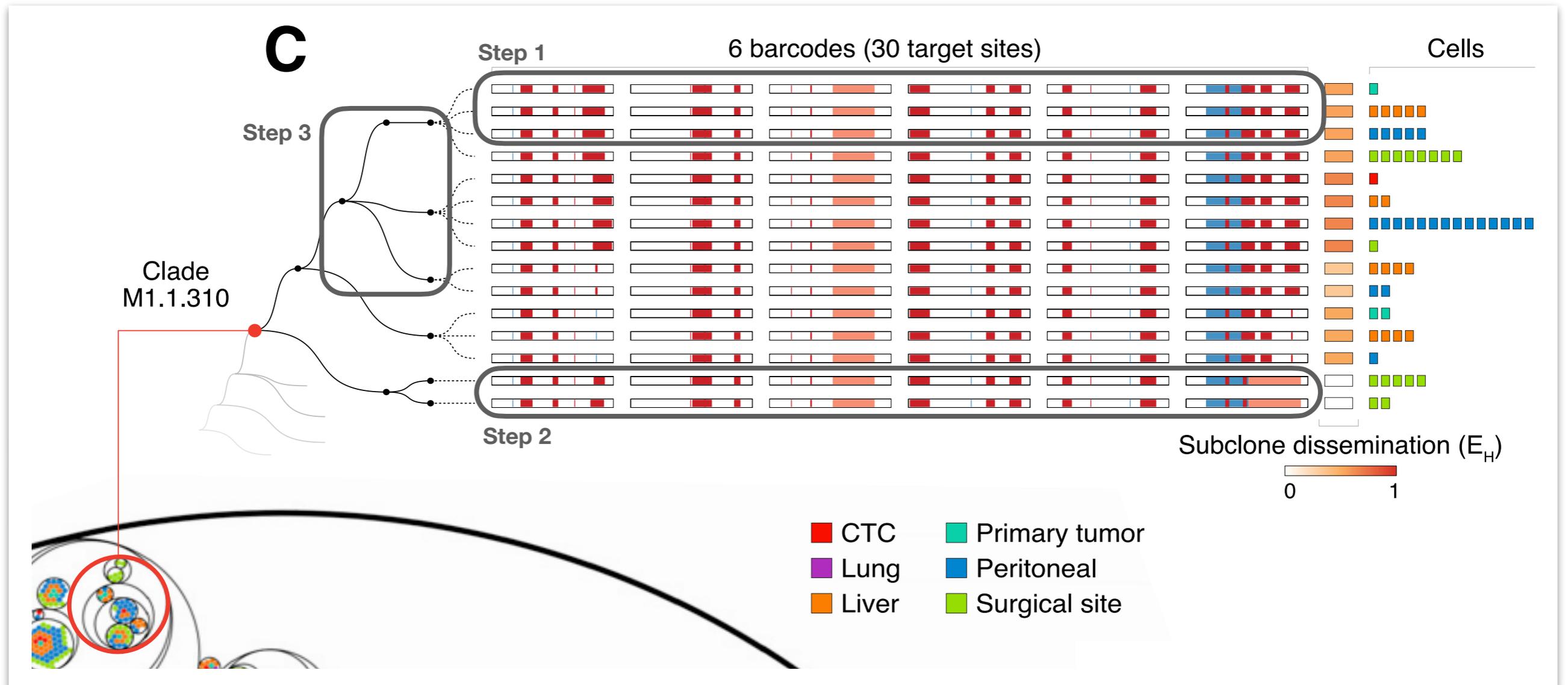


Recovered on average:

- **Mouse 1:** 18.5 target sites (3.7 barcodes) / cell
- **Mouse 2:** 8.5 target sites (1.7 barcodes) / cell



High-res subclonal lineage reconstruction

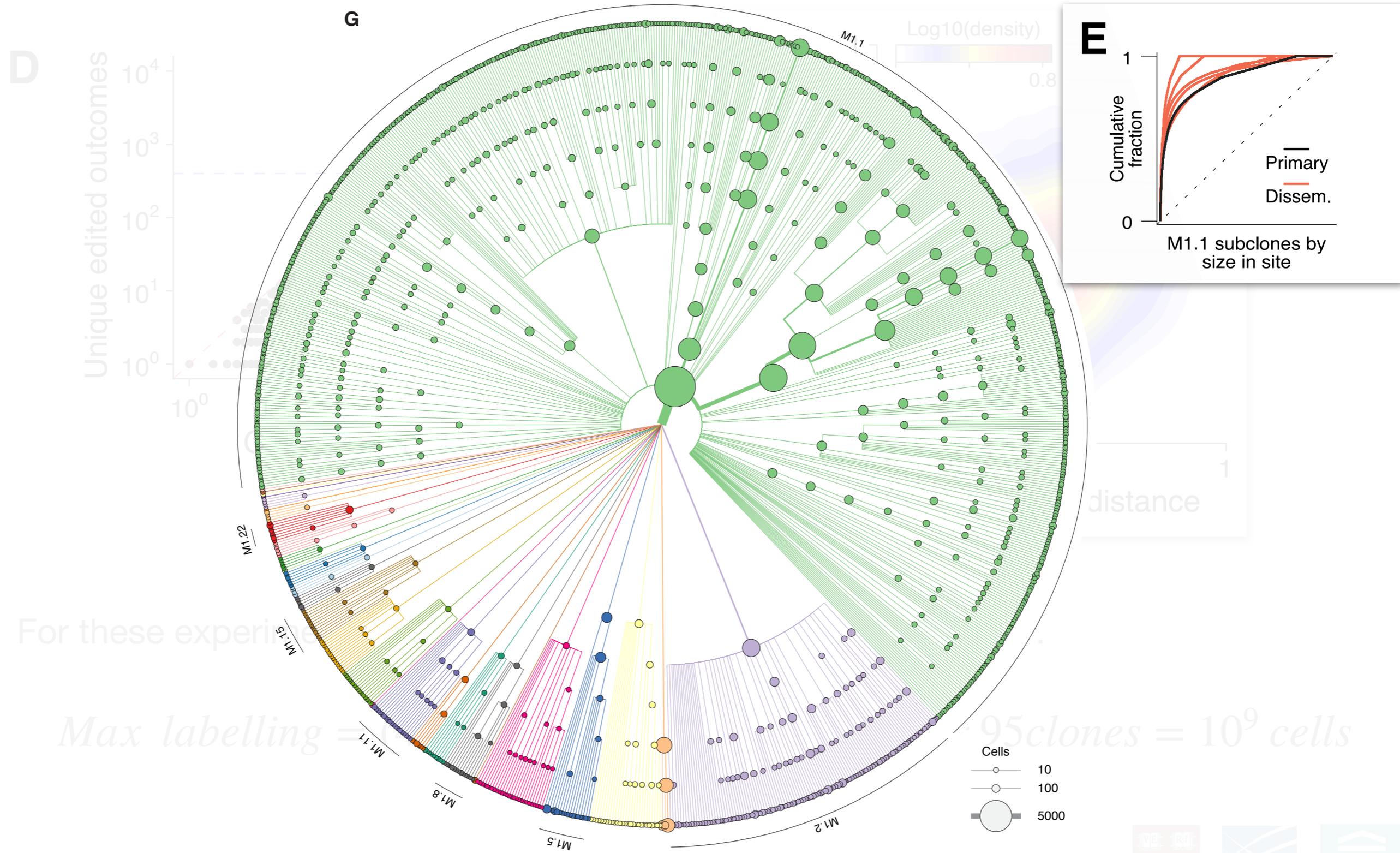


Step 1: assemble “barcode of barcodes”

Step 2: group cells into subclones

Step 3: reconstruct phylogenetic relationships between subclones

High-res subclonal lineage reconstruction

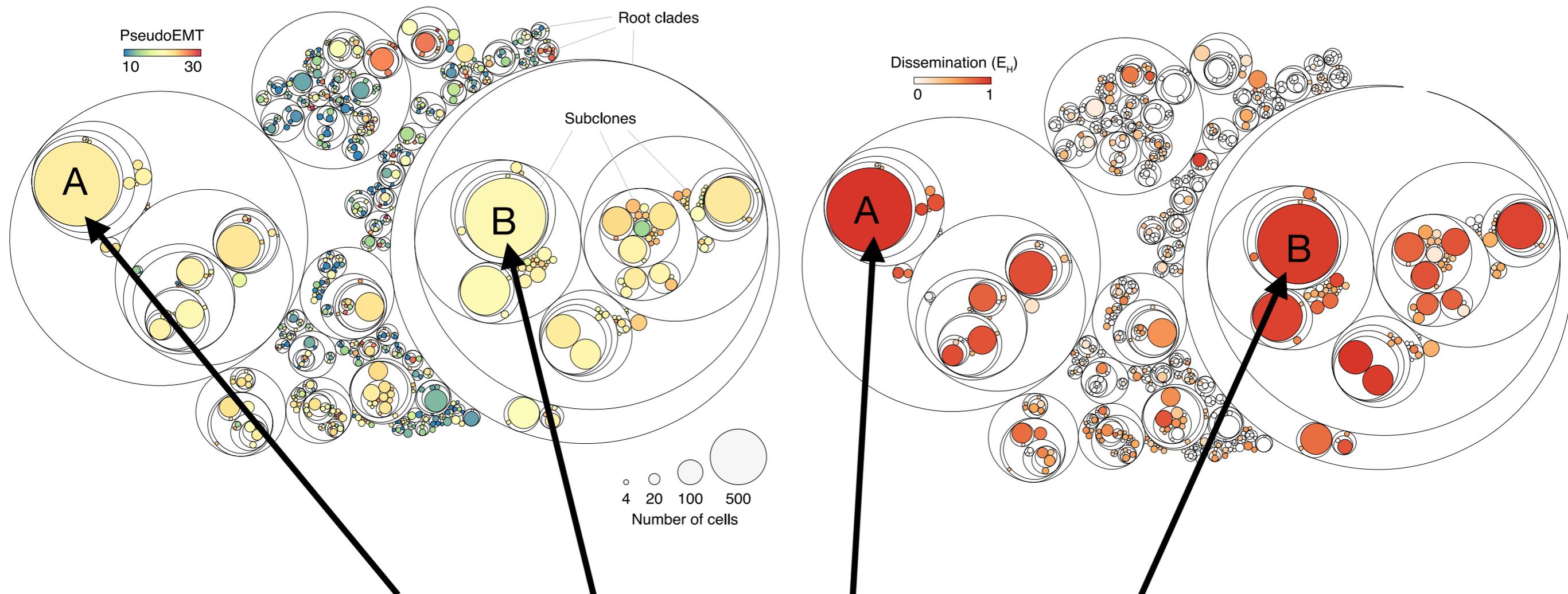


From Figures 4, S6 of Simeonov et al., Cancer Cell (2021)



Peak metastatic aggression corresponds to late-hybrid EMT states

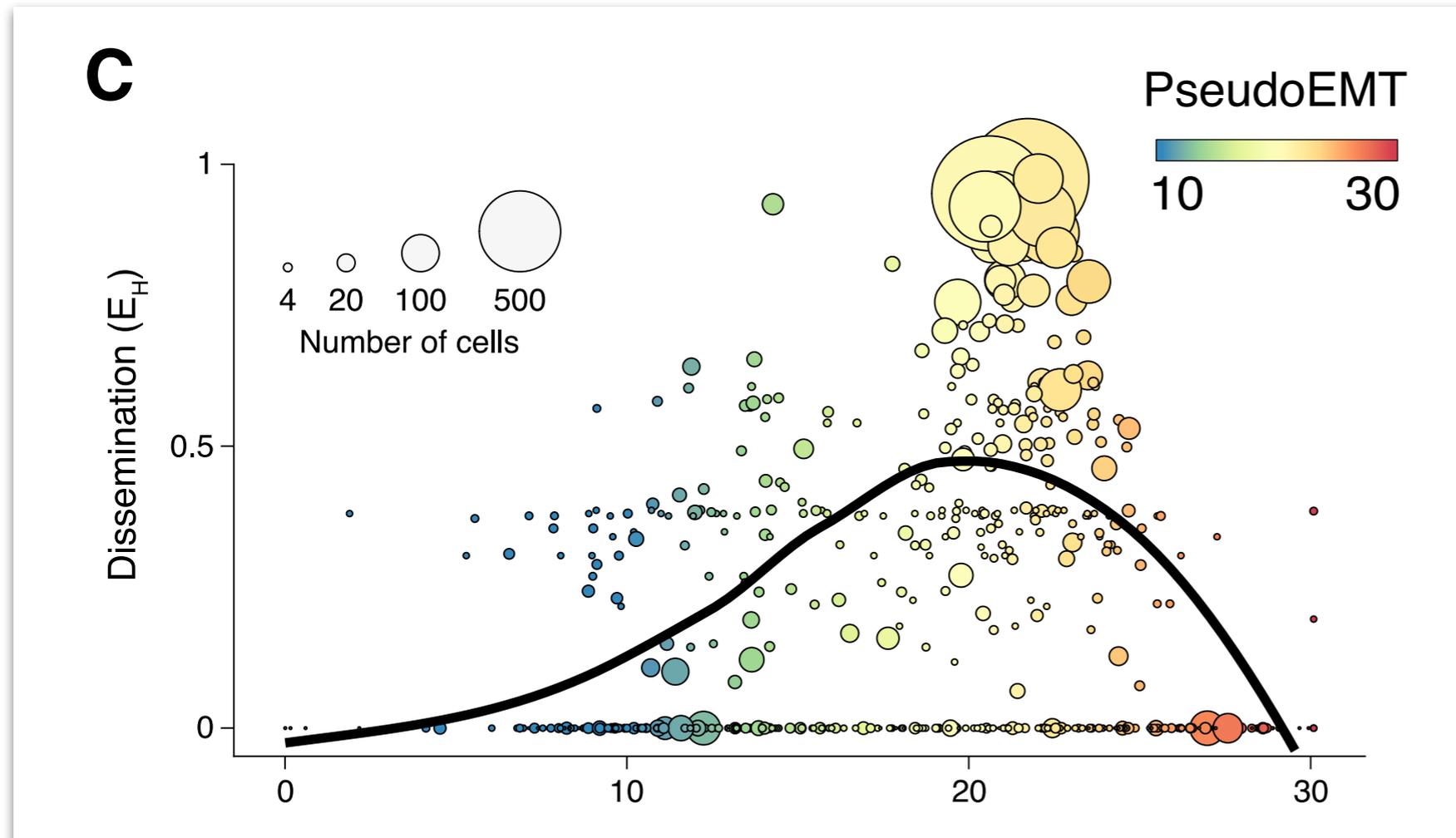
How does the range of **intraclonal EMT states** relate **to subclonal behavior**?



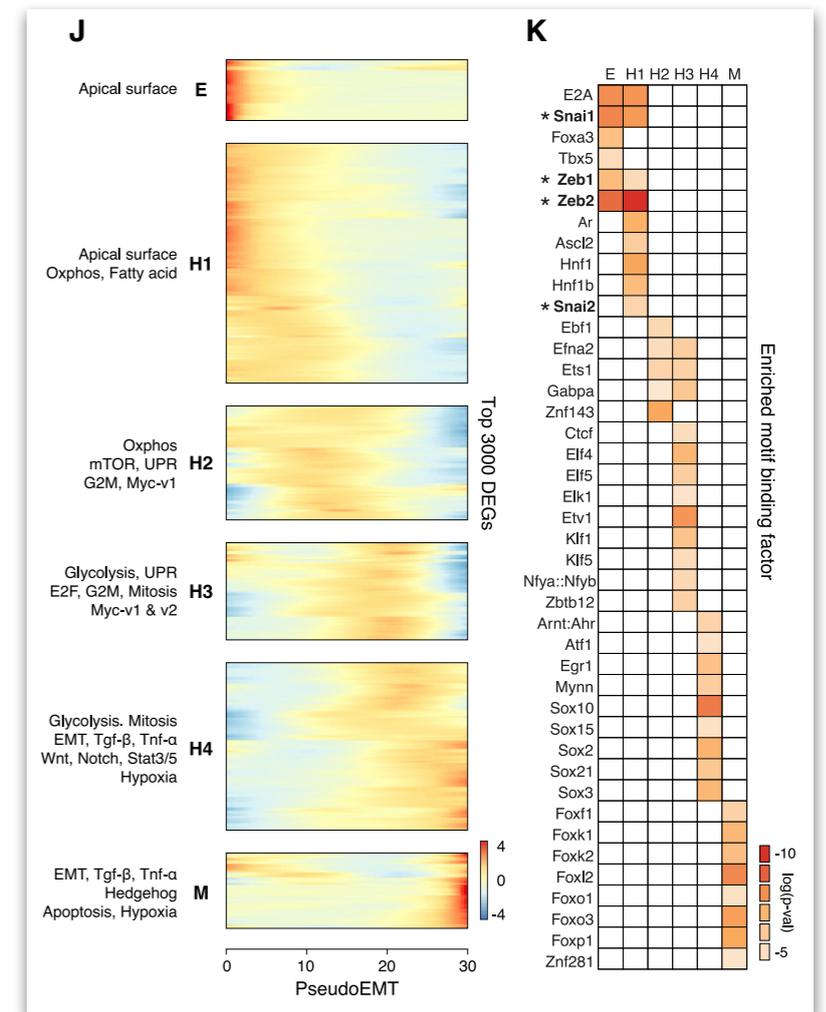
Direct evidence that **hybrid EMT states are more metastatic than EMT extremes**



Peak metastatic aggression corresponds to late-hybrid EMT states



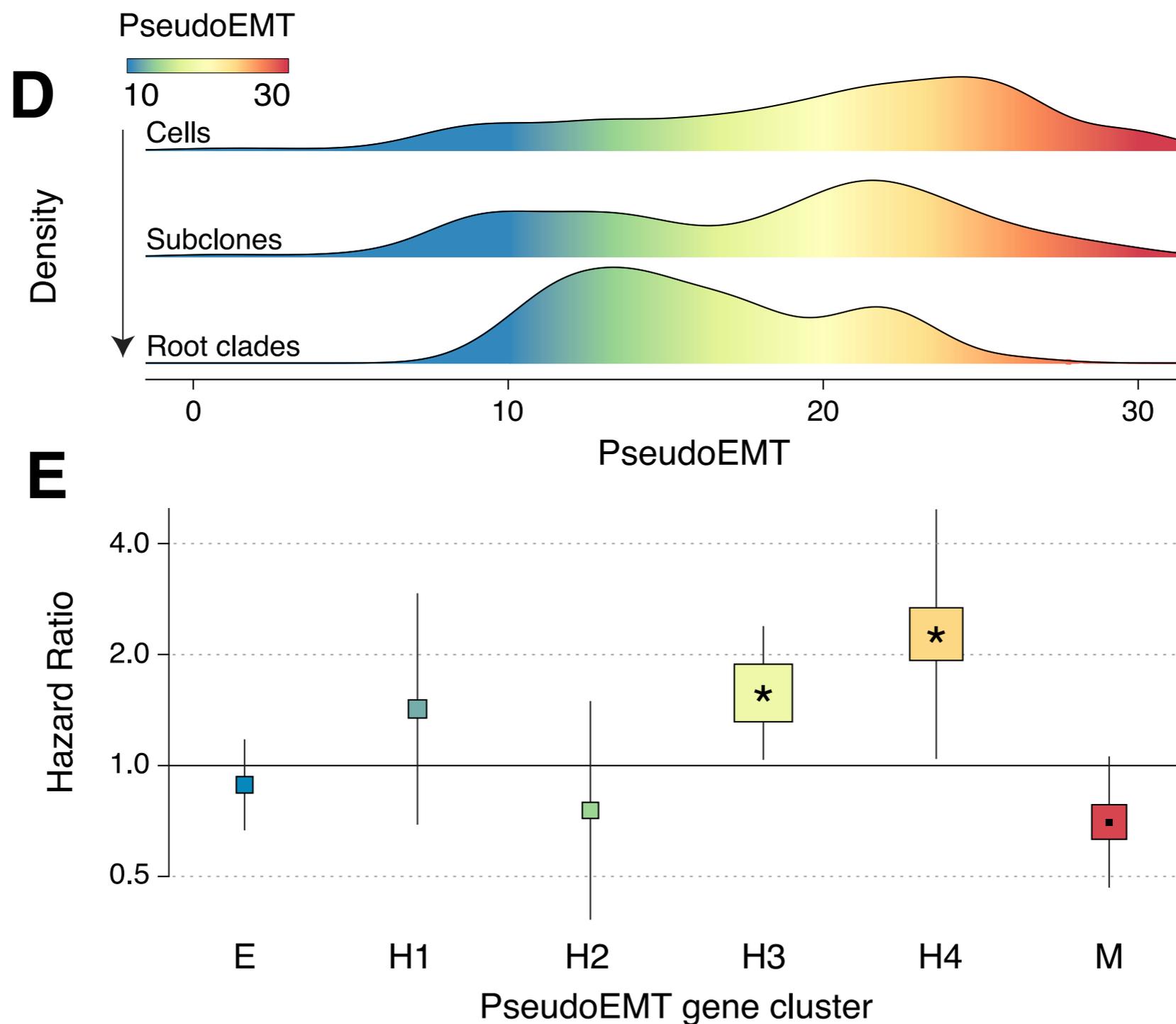
Remember from Figure 3....



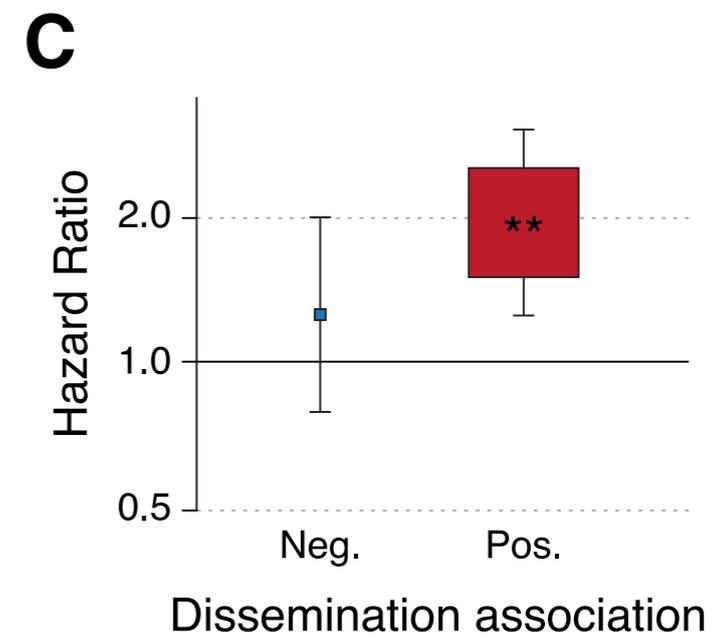
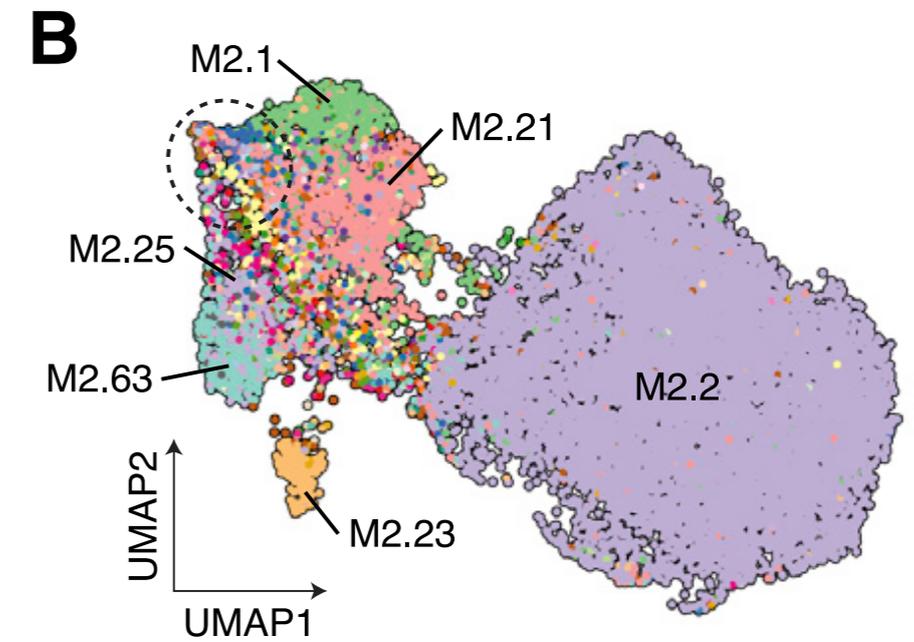
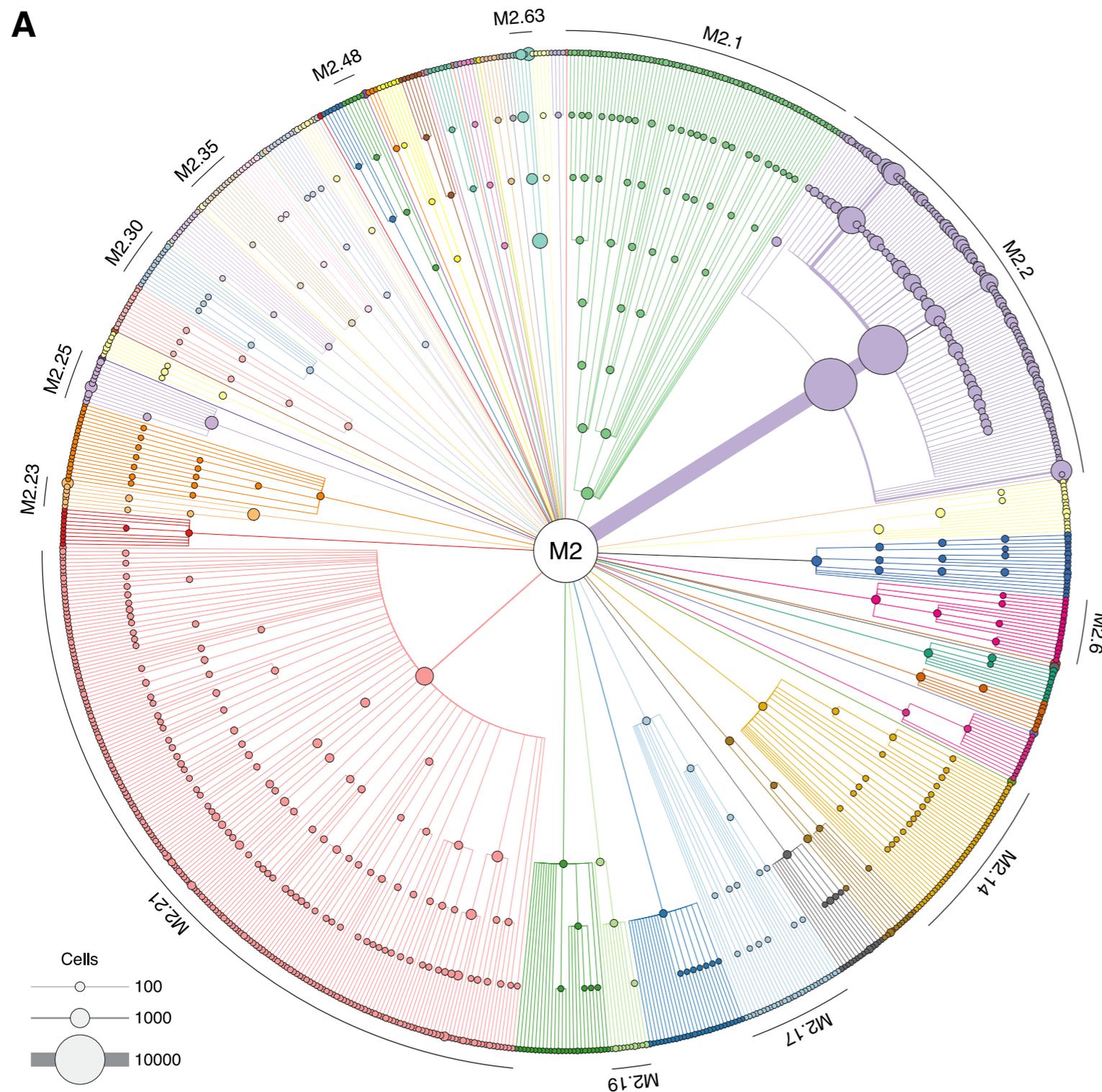
Important theme: this paper provides a lot of evidence that EMT, while its own process is very inherent to cancer metastasis - remember we're studying a cancer model not an EMT model



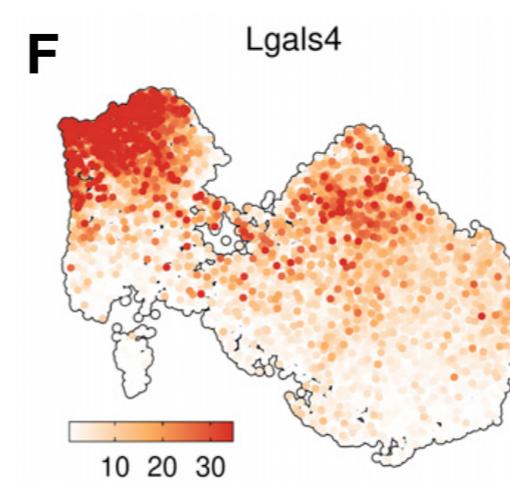
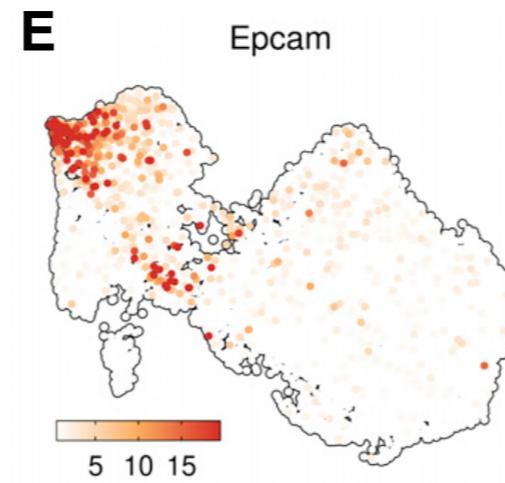
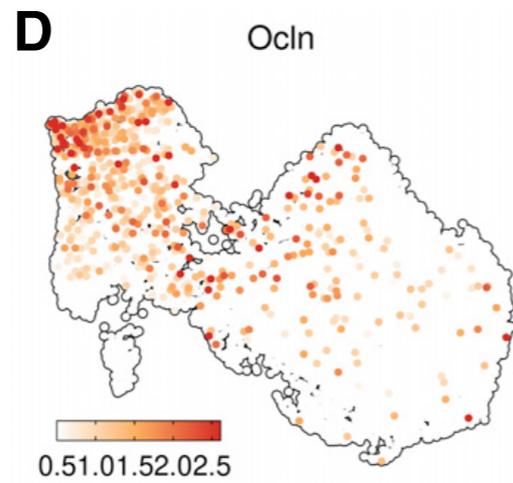
Peak metastatic aggression corresponds to late-hybrid EMT states



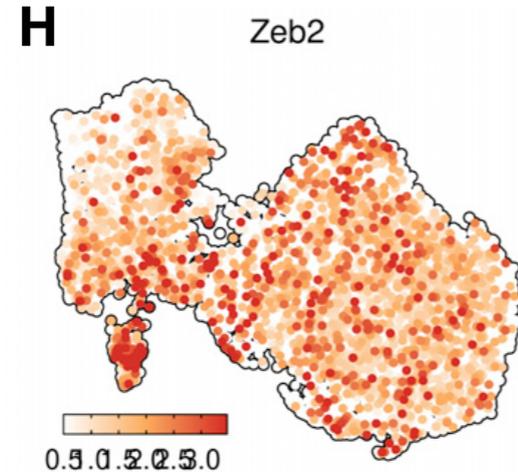
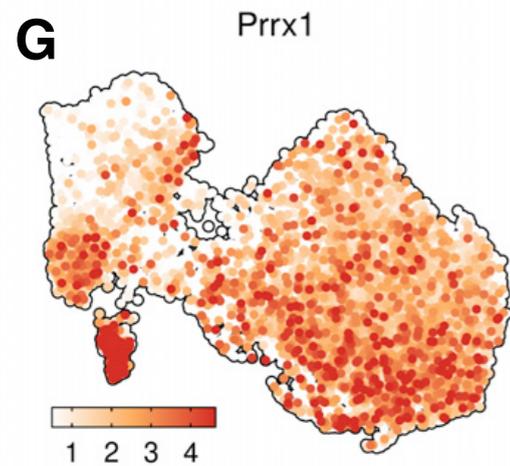
Complementary process to canonical EMT



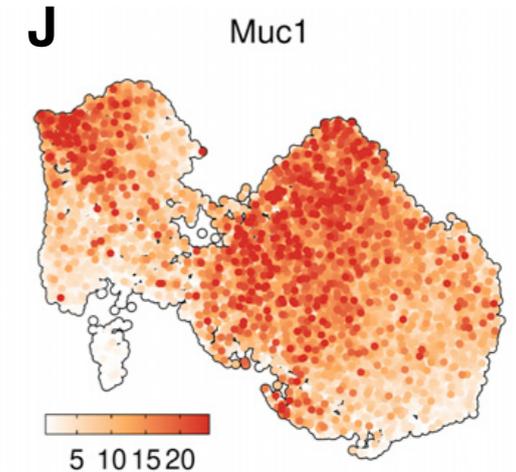
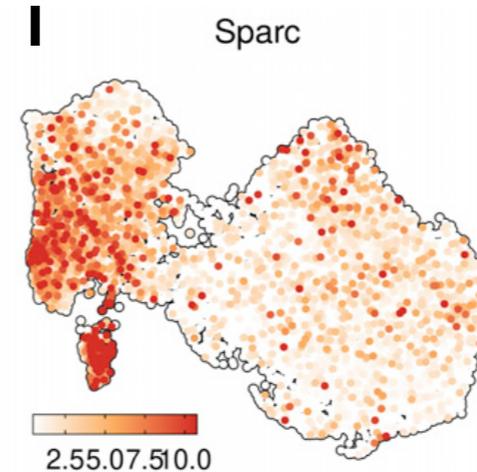
Complementary process to canonical EMT



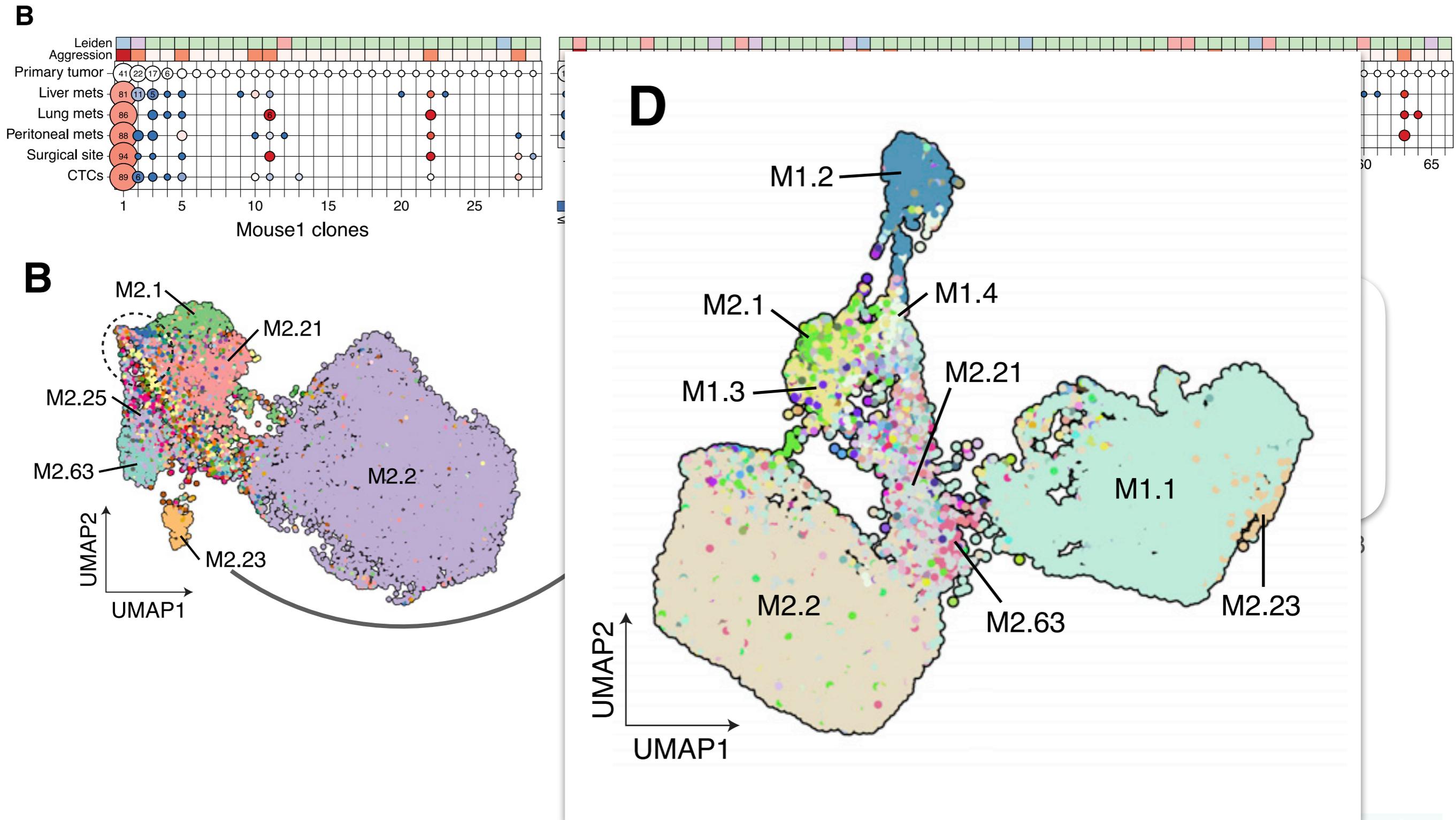
GEX as expected



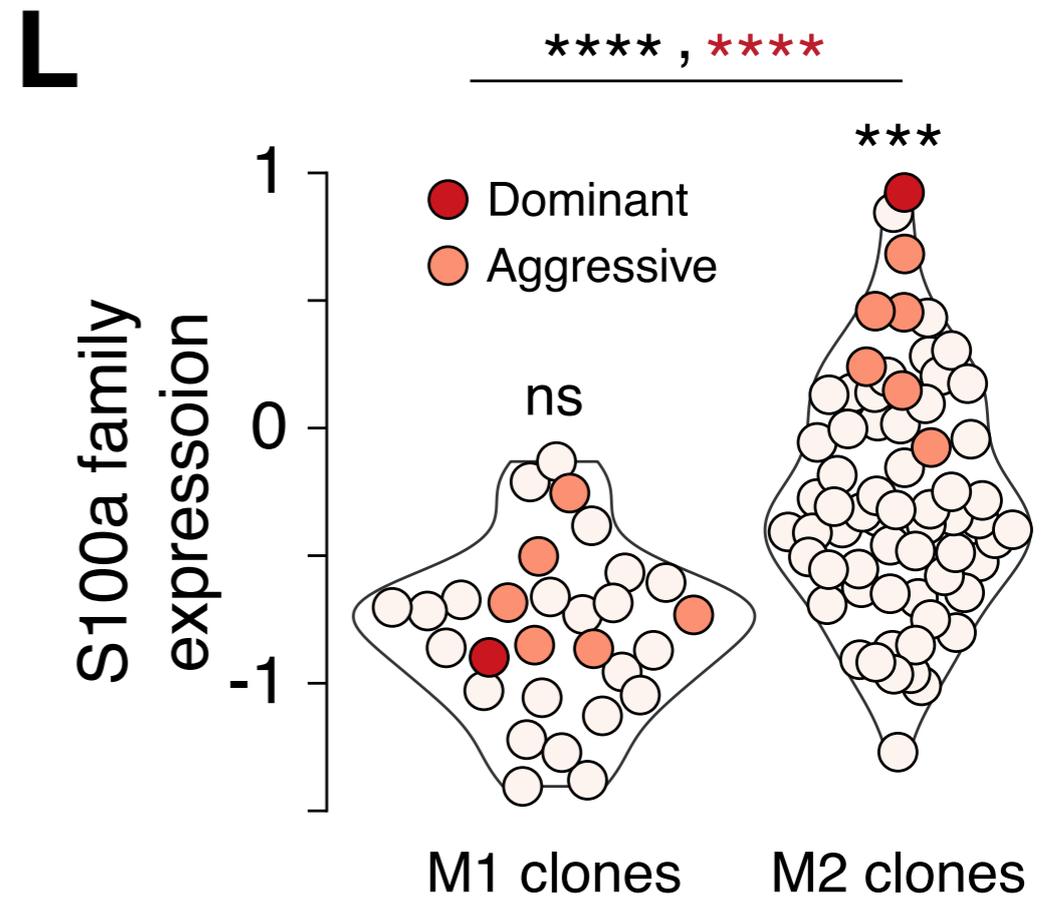
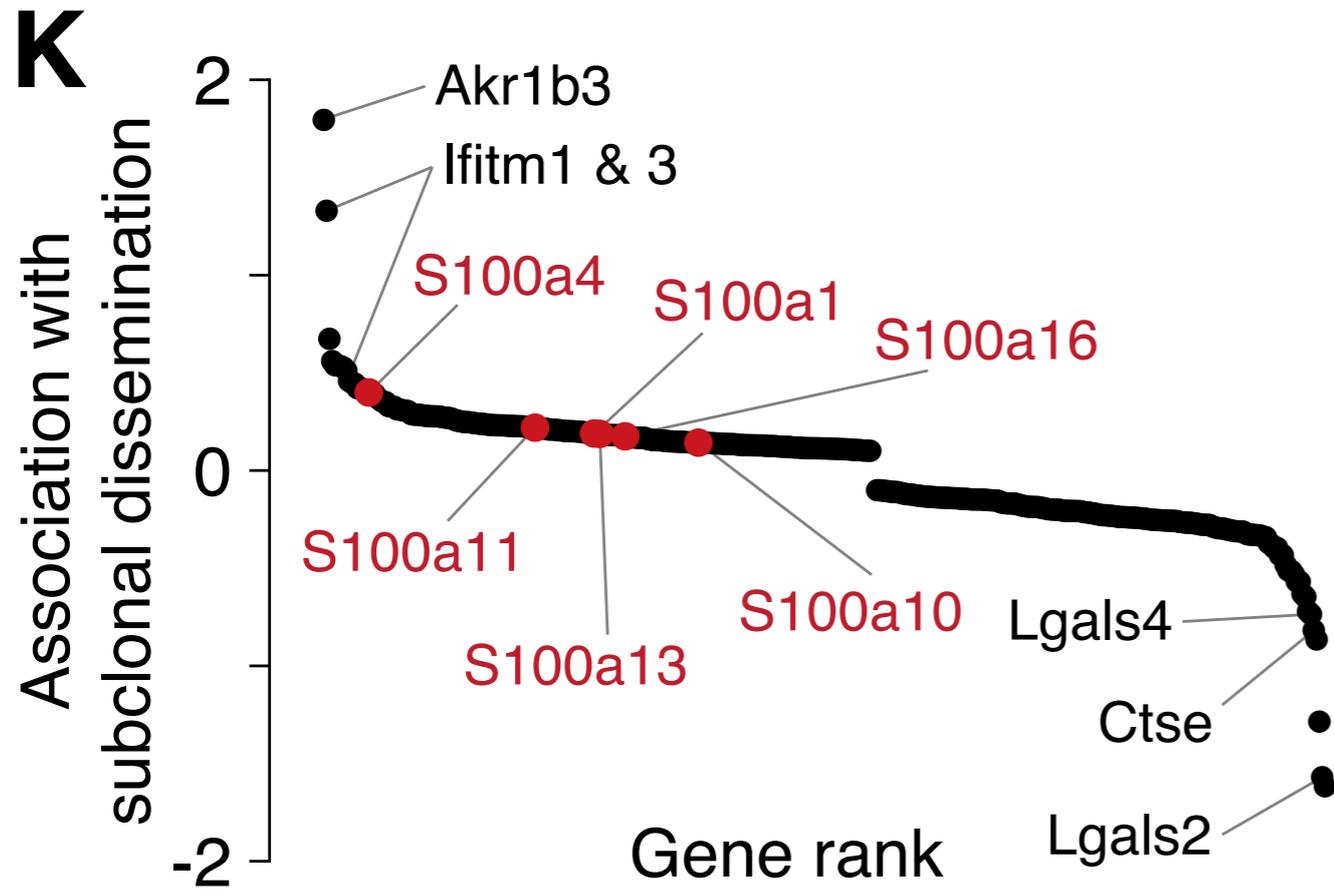
GEX contradictory



Complementary process to canonical EMT

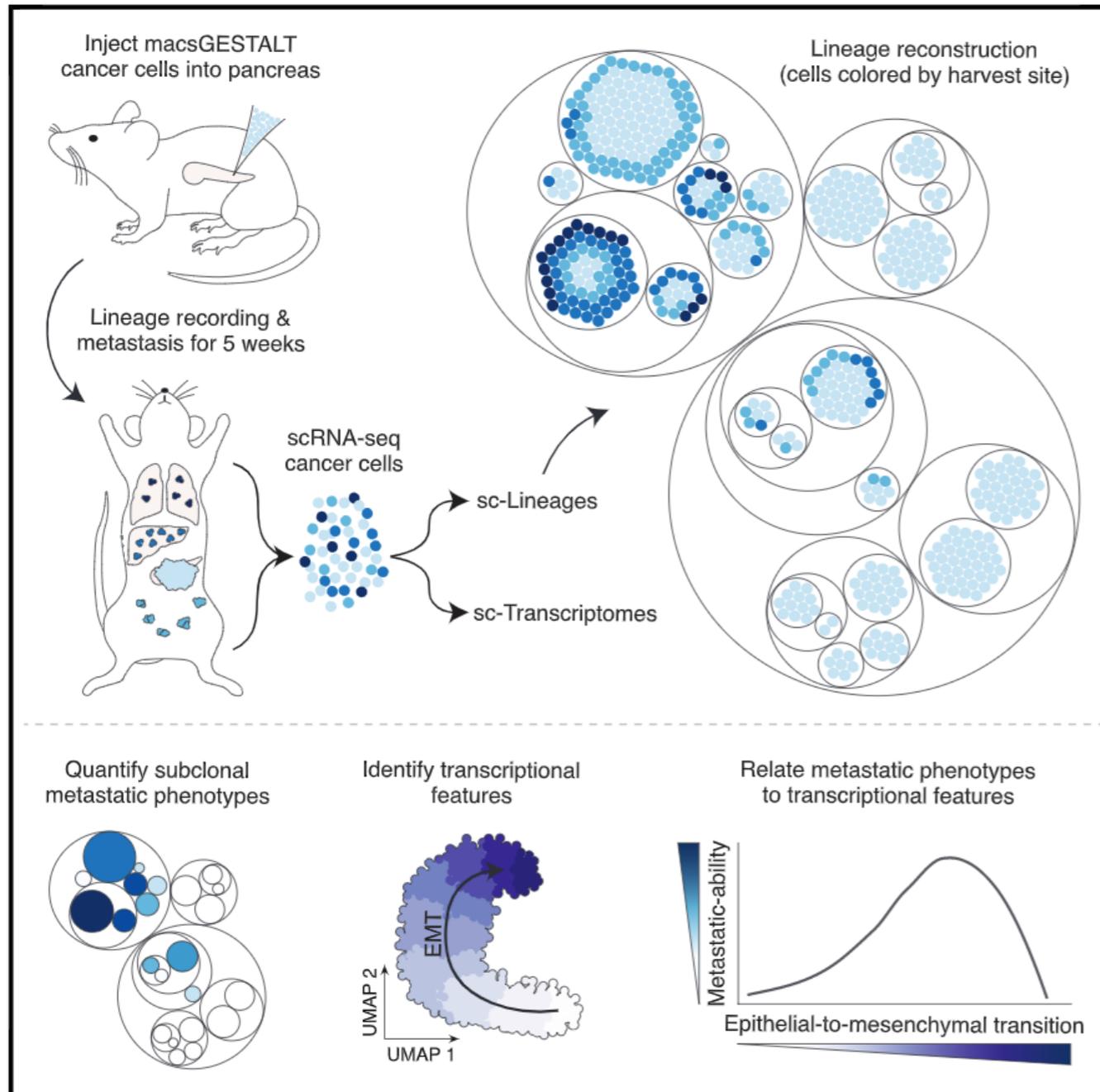


Complementary process to canonical EMT



Major contributions of macsGESTALT

Graphical abstract



- macsGESTALT is an inducible lineage-recorder
- In vivo model of pancreatic cancer metastasis
- Finding recurrent drivers across cancers remains elusive
- Metastatically competent model wherein most clones do NOT metastasize
- In EMT, metastatic aggression rises and peaks during a late intermediate hybrid stage
- ID gene sets within EMT hybrid stages that are predictive of human survival outcome (in 2 cancers, not in 3)
- *S100* genes were found across metastatic subpopulations

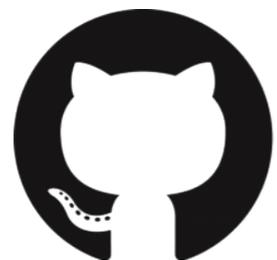


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